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 71) Applicant: BAYER CORPORATION [US/US]; 10 Road, Pittsburgh, PA 15205 (US). 72) Inventors: HU, Yinghe; 17 Falcon Crest Drive, Nort CT 06473 (US). McCALEB, Michael, L.; 447 Drive, Madison, CT 06443 (US). BLOOMQUIST.; 405 Stevenson Road, New Haven, CT 065 FLORES-RIVEROS, Jaime, R.; 27 Carmel Court, CT 06443 (US). CORNFIELD, Linda, J.; 3 Hidde Road, Hamden, CT 06518 (US). 74) Agents: GREENMAN, Jeffrey, M. et al.; Bayer Cor 400 Morgan Lane, West Haven, CT 06516 (US). 	h Have 7 Bartle T, Bria 15 (US Madiso en Broo	Without international search report and to be republished upon receipt of that report.

(57) Abstract

The present invention provides novel NPY/PYY receptor proteins and the nucleic acid sequence encoding them. The invention is directed to the isolation, characterization, and pharmacological use of these receptors and nucleic acids. In particular, this invention provides human and rat NPY/PYY receptors (which we call the NPY Y5 receptor) and nucleic acids. Also provided are recombinant expression constructs useful for transfecting cells and expressing the protein in vitro and in vivo. The invention further provides methods for detecting expression levels of the protein as well as methods for screening for receptor antagonists and agonists to be used for the treatment of obesity or anorexia, respectively.

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NEUROPEPTIDE Y RECEPTOR Y5 AND NUCLEIC ACID SEQUENCE BACKGROUND OF THE INVENTION

Field of the Invention

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This invention relates to a novel neurotransmitter Neuropeptide Y receptor, its nucleic acid sequence, and compounds, compositions, and methods for their use.

Summary of the Related Art

Neuropeptide Y (NPY) is a 36-amino acid peptide neurotransmitter that is located throughout the central and peripheral nervous systems. Tatemoto, *Proc. Natl. Acad. Sci. USA* 79, 5485 (1982); Hazlewood, *Proc. Soc. Exp. Biol. Med.* 202, 44 (1993). It affects a broad range of phenomena, including blood pressure regulation, memory, anxiolysis/sedation, food and water appetite, vascular and other smooth muscle activity, intestinal electrolyte secretion, and urinary sodium excretion. *E.g.*, Colmers and Wahlestedt, *The Biology of Neuropeptide Y and Related Peptides* (Humana Press, Totowa, NJ, 1993); Kalra et al., *Phys. & Behavior* 50, 5 (1991).

Peptide YY (PYY) is also a 36 amino acid peptide and has significant sequence homology (70%) to NPY. Tatemoto et al., *Nature* 296, 659 (1982). Its anatomical distribution is similar to that of NPY, although it is located mainly in the endocrine cells of the lower gastrointestinal tract. Bottcher et al., *Regul. Pept.* 8, 261 (1984). Like NPY, PYY stimulates feeding in rats. Morley et al., *Brain Res.* 341, 200 (1985). Along with the pancreatic polypeptide (PP), NPY and PYY have a common tertiary structure, characterized by the so-called PP-fold. Glover, *Eur. J. Biochem.* 142, 379 (1985). Both NPY and PYY show about a 50% sequence homology with PP.

Because of their structural similarities, NPY and PYY have a number of common receptors. At least four receptor subtypes, Y1, Y2, Y3, and Y4/PP, have been identified. The affinity for NPY, PYY, and various fragments thereof varies among the subtypes. See, e.g., Bard et al. (WO 95/17906) and references cited therein. For example, Y1 and Y2 subtypes have high affinity for NPY and PYY. Whereas Y1 has high affinity for (Leu³¹Pro³⁴)NPY ((LP)NPY)and low affinity for (13-36)NPY, Y2 behaves oppositely. Y3 has high affinity for NPY but low affinity for PYY. Y4/PP has a high affinity for PP but relatively low affinity for NPY.

Wahlestedt (WO 93/24515) and Larhammar et al. (J. Biol. Chem. 267, 10935 (1992)) describe the cloning and identification of the human Y1-type NPY/PYY receptor isolated from

human fetal brain tissue. Selbie et al. (WO 93/09227) disclosed the full length cDNA sequence of the Y1 receptor from human hippocampus. Eva et al. (FEBS Lett. 271, 81 (1990)) cloned the NPY Y1 receptor from rat forebrain. Eva et al. (FEBS Lett. 314, 285 (1992)) cloned the NPY Y1 receptor from murine genomic DNA.

The Y2-type receptor has also been cloned. Gerald et al. (WO 95/21245) disclosed the cDNA sequence of human hippocampal Y2 and two rat Y2 clones. Rose et al. (*J. Biol. Chem.* 270, 22661 (1995)) disclosed the cDNA sequence of the Y2 receptor from a human neuroblastoma cell line.

Bard et al. (supra) and Lundell et al. (J. Biol. Chem. 270, 29123 (1995)) described cloning the cDNA sequence of the Y4/PP receptor from both rat spleen and human placenta.

To date, the Y3 receptor has not been cloned.

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Because of the important role of NPY and PYY in a number of physiological processes, such as feeding, there is a strong need to further develop materials and methods for investigating the mechanistic behavior of these compounds and for treating diseased and other abnormal states associated with the physiological processes in which NPY and PYY act. Specifically, the NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, do not bind to the previously identified NPY/PYY receptors with affinities consistent with the feeding response. Accordingly, there is a need and desire to identify the NPY/PYY receptor that is responsible for the feeding response. Antagonists to such a receptor could be used to treat obesity and diabetes by reducing appetite and food consumption.

SUMMARY OF THE INVENTION

The present invention provides, *inter alia*, novel NPY/PYY receptor proteins. Also provided are the nucleic acid sequences encoding these novel receptor proteins, as well as compounds and methods for using these proteins and their nucleic acid sequences.

The present invention provides novel proteins, nucleic acids, and methods useful for developing and identifying compounds for the treatment of such diseases and disorders as obesity. Identified and disclosed herein is the protein sequence for a novel receptor for the neurotransmitters Neuropeptide Y (NPY) and Peptide YY (PYY) and the nucleic acid sequence encoding this receptor, which we call the NPY Y5 (or simply "Y5") receptor. The importance of this discovery is manifested in the effects of NPY, which include blood pressure regulation, memory enhancement, anxiolysis/sedation, and increased food intake. Thus, this receptor

protein is useful for screening for NPY/PYY agonist and antagonist activity for controlling these conditions.

In one aspect of the present invention, we provide isolated nucleic acid sequences for a novel NPY and PYY receptor, the Y5 receptor. In particular, we provide the cDNA sequences encoding for the rat and human receptors and isoforms thereof. These nucleic acid sequences have a variety of uses. For example, they are useful for making vectors and for transforming cells, both of which are ultimately useful for production of the Y5 receptor protein. They are also useful as scientific research tools for developing nucleic acid probes for determining receptor expression levels, e.g., to identify diseased or otherwise abnormal states. They are useful for developing analytical tools such as antisense oligonucleotides for selectively inhibiting expression of the receptor gene to determine physiological responses.

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In another aspect of the present invention, we provide a homogenous composition comprising the receptor Y5 protein. The protein is useful for screening drugs for agonist and antagonist activity, and, therefore, for screening for drugs useful in regulating physiological responses associated with the Y5 receptor. Specifically, antagonists to the Y5 receptor could be used to treat obesity and diabetes by reducing appetite and food consumption, whereas agonists could be used for the treatment of anorexic conditions. The proteins are also useful for developing antibodies for detection of the protein.

Flowing from the foregoing are a number of other aspects of the invention, including (a) vectors, such as plasmids, comprising the receptor Y5 nucleic acid sequence that may further comprise additional regulatory elements, e.g., promoters, (b) transformed cells that express the Y5 receptor, (c) nucleic acid probes, (d) antisense oligonucleotides, (e) agonists, (f) antagonists, and (g) transgenic mammals. Further aspects of the invention comprise methods for making and using the foregoing compounds and compositions.

The foregoing merely summarizes certain aspects of the present invention and is not intended, nor should it be construed, to limit the invention in any manner. All patents and other publications recited herein are hereby incorporated by reference in their entirety.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 displays the competition curves of various peptides for [125I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells.

Figure 2 displays saturation curves for specific binding of [125I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells.

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DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention comprises, in part, a novel NPY/PYY receptor protein, the Y5 receptor. Particularly preferred embodiments of the Y5 receptor are those having an amino acid sequence substantially the same as SEQ ID NOs 2, 4, or 6. As used herein, reference to the Y5 receptor is meant as a reference to any protein having an amino acid sequence substantially the same as SEQ ID NOs 2, 4, or 6. The present invention also comprises the nucleic acid sequence encoding the Y5 protein, which nucleic acid sequences is substantially the same as SEQ ID NOs 1, 3, or 5. Receptors SEQ ID NOs 2 and SEQ ID NO 4 are rat Y5 receptors and appear to be allelic variations, with SEQ ID NO 4 the most commonly occurring and, therefore, the preferred embodiment of the rat Y5 receptor of this invention. SEQ ID NO 6 is the human Y5 receptor and its preferred embodiment.

As used herein, a protein "having an amino acid sequence substantially the same as SEQ ID NO x" (where "x" is the number of one of the protein sequences recited in the Sequence Listing) means a protein whose amino acid sequence is the same as SEQ ID NO x or differs only in a way such that IC₅₀[(3-36)NPY], IC₅₀[(Leu³¹Pro³⁴)NPY], and IC₅₀[(Leu³¹Pro³⁴)(3-36)NPY] as determined according to the method detailed in Example 4, *infra*, are less than or equal to 30 nM. The NPY fragments (3-36)NPY, (Leu³¹Pro³⁴)NPY and (Leu³¹Pro³⁴)(3-36)NPY induce a feeding response. Those skilled in the art will appreciate that conservative substitutions of amino acids can be made without significantly diminishing the protein's affinity for NPY, PYY, and fragments and analogs thereof. Other substitutions may be made that increase the protein's affinity for these compounds. Making and identifying such proteins is a routine matter given the teachings herein, and can be accomplished, for example, by altering the nucleic acid sequence encoding the protein (as disclosed herein), inserting it into a vector, transforming a cell, expressing the nucleic acid sequence, and measuring the binding affinity of the resulting protein, all as taught herein.

As used herein the term "a molecule having a nucleotide sequence substantially the same as SEQ ID NO y" (wherein "y" is the number of one of the protein-encoding nucleotide sequences listed in the Sequence Listing) means a nucleic acid encoding a protein "having an amino acid sequence substantially the same as SEQ ID NO y+1" (wherein "y+1" is the number of the amino acid sequence for which nucleotide sequence "y" codes) as defined above. This definition is intended to encompass natural allelic variations in the Y5 sequence. Cloned nucleic acid provided by the present invention may encode Y5 protein of any species of origin, including (but not limited to), for example, mouse, rat, rabbit, cat, dog, primate, and human. Preferably the nucleic acid provided by the invention encodes Y5 receptors of mammalian, and most preferably, rat or human origin.

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The invention also includes nucleotide sequences encoding chimeric proteins comprised of parts of the Y5 receptor and parts of other related seven-transmembrane receptors.

The 6B clone (SEQ ID NO 1) (see Example 2, infra) has a 2.4 kb cDNA insert with a open reading frame from nucleotide 248 to 1582 that encodes a 445 amino acid protein (SEQ ID NO 2). Hydrophobicity plot analysis using PEPPLOT of GCG shows that the Y5 receptor has seven transmembrane-like domains, indicating it might be a G-protein-coupled receptor. Unlike other known subtypes of NPY receptor family, the third intracellular loop of the Y5 receptor is unusually long. Another novel feature of the Y5 peptide sequence is that it has a much shorter C-terminal tail sequence than other known members of the NPY receptor family. It is also important to note that the Y5 sequence shows only 30-33% amino acid sequence identity to other NPY receptors.

Nucleic acid hybridization probes provided by the invention are DNAs consisting essentially of the nucleotide sequences complementary to any sequence depicted in SEQ ID NOs 1, 3, and 5 that is effective in nucleic acid hybridization. Nucleic acid probes are useful for detecting Y5 gene expression in cells and tissues using techniques well-known in the art, including, but not limited to, Northern blot hybridization, in situ hybridization, and Southern hybridization to reverse transcriptase - polymerase chain reaction product DNAs. The probes provided by the present invention, including oligonucleotide probes derived therefrom, are also useful for Southern hybridization of mammalian, preferably human, genomic DNA for screening for restriction fragment length polymorphism (RFLP) associated with certain genetic disorders. As used herein, the term complementary means a nucleic acid having a sequence that is sufficiently complementary in the Watson-Crick sense to a target nucleic acid to bind to the

target under physiological conditions or experimental conditions those skilled in the art routinely use when employing probes.

Receptor Y5 binds various fragments and analogs of NPY and PYY with affinities different from that of the known receptors. The rank order of binding affinity of receptor Y5 was found to be:

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NPY = (LP)NPY = PYY = (3-36)NPY = (LP)(3-36)NPY > (10-36)NPY > (18-36)NPY Table 1, *infra*, presents a more detailed affinity profile of the Y5 receptor for NPY, PYY, and various fragments thereof. As used herein, a protein having substantially the same affinity profile as the Y5 receptor means a protein in which the IC_{50} of each of the peptides listed in Table 1, *infra*, is no more than an order of magnitude greater than those listed in Table 1 for each of the respective peptides as measured according to the methods described in Example 4. Importantly, the NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, do not bind to the previously identified NPY/PYY receptors with affinities consistent with the feeding response.

The production of proteins such as receptor Y5 from cloned genes by genetic engineering means is well known in this art. The discussion which follows is accordingly intended as an overview of this field, and is not intended to reflect the full state of the art.

DNA which encodes receptor Y5 may be obtained, in view of the instant disclosure, by chemical synthesis, by screening reverse transcripts of mRNA from appropriate cells or cell line cultures, by screening genomic libraries from appropriate cells, or by combinations of these procedures, as illustrated below. Screening of mRNA or genomic DNA may be carried out with oligonucleotide probes generated from the Y5 gene sequence information provided herein. Probes may be labeled with a detectable group such as a fluorescent group, a radioactive atom or a chemiluminescent group in accordance with known procedures and used in conventional hybridization assays, as described in greater detail in the Examples below. In the alternative, the Y5 gene sequence may be obtained by use of the polymerase chain reaction (PCR) procedure, with the PCR oligonucleotide primers being produced from the Y5 gene sequence provided herein. See U.S. Patent Nos. 4,683,195 to Mullis et al. and 4,683,202 to Mullis.

Receptor Y5 may be synthesized in host cells transformed with a recombinant expression construct comprising a nucleic acid encoding the receptor Y5. Such a recombinant expression construct can also be comprised of a vector that is a replicable DNA construct. Vectors are used herein either to amplify DNA encoding Y5 and/or to express DNA which

encodes Y5. For the purposes of this invention, a recombinant expression construct is a replicable DNA construct in which a DNA sequence encoding Y5 is operably linked to suitable control sequences capable of effecting the expression of Y5 in a suitable host. The need for such control sequences will vary depending upon the host selected and the transformation method chosen. Generally, control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites, and sequences which control the termination of transcription and translation. Amplification vectors do not require expression control domains. All that is needed is the ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants. See, Sambrook et al., Molecular Cloning: A Laboratory Manual (2nd Edition, Cold Spring Harbor Press, New York, 1989).

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Vectors useful for practicing the present invention include plasmids, viruses (including phage), retroviruses, and integratable DNA fragments (*i.e.*, fragments integratable into the host genome by homologous recombination). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself. Suitable vectors will contain replicon and control sequences which are derived from species compatible with the intended expression host. The vectors may be self-replicating. Suitable vectors for the purposes of the present invention include pBluescript, pcDNA3, and, for insect cells, baculovirus. A preferred vector is the plasmid pcDNA3 (Invitrogen).

Construction of suitable vectors containing the desired coding and control sequences employs standard ligation and restriction techniques that are well understood in the art. Isolated plasmids, DNA sequences, or synthesized oligonucleotides are cleaved, tailored, and relegated in the form desired.

Site-specific DNA cleavage is performed by treating with the suitable restriction enzyme (or enzymes) under conditions that are generally understood in the art, and the particulars of which are specified by the manufacturer of these commercially available restriction enzymes. See, e.g., New England Biolabs, Product Catalog. In general, about 1 µg of plasmid or DNA sequence is cleaved by one unit of enzyme in about 20 µl of buffer solution. Often excess of restriction enzyme is used to ensure complete digestion of the DNA substrate. Incubation times of about one hour to two hours at about 37°C are workable, although variations are tolerable. After each incubation, protein is removed by extraction with phenol/chloroform, and may be followed by ether extraction. The nucleic acid may be recovered from aqueous fractions by

precipitation with ethanol. If desired, size separation of the cleaved fragments may be performed by polyacrylamide gel or agarose gel electrophoresis using standard techniques. A general description of size separations is found in *Methods in Enzymology* 65, 499-560 (1980).

Transformed host cells are cells which have been transformed or transfected with recombinant expression constructs made using recombinant DNA techniques and comprising mammalian Y5-encoding sequences. Preferred host cells for transient transfection are COS-7 cells. Transformed host cells may ordinarily express Y5, but host cells transformed for purposes of cloning or amplifying nucleic acid hybridization probe DNA need not express the receptor. When expressed, the mammalian Y5 protein will typically be located in the host cell membrane. See, Sambrook et al., ibid.

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Cultures of cells derived from multicellular organisms are desirable hosts for recombinant Y5 protein synthesis. In principal, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. However, mammalian cells are preferred, as illustrated in the Examples. Propagation of such cells in cell culture has become a routine procedure. See Tissue Culture (Academic Press, Kruse & Patterson, Eds., 1973). Examples of useful host cell lines are bacteria cells, insect cells, yeast cells, human 293 cells, VERO and HeLa cells, LMTK cells, and WI138, BHK, COS-7, CV, and MDCK cell lines. Human 293 cells are preferred.

The invention provides homogeneous compositions of mammalian Y5 produced by transformed eukaryotic cells as provided herein. Such homogeneous compositions are intended to be comprised of mammalian Y5 protein that comprises at least 90% of the protein in such homogeneous composition. The invention also provides membrane preparation from cells expressing Y5 as the result of transformation with a recombinant expression construct, as described here.

Mammalian Y5 protein made from cloned genes in accordance with the present invention may be used for screening compounds for Y5 agonist or antagonist activity, or for determining the amount of a Y5 agonist or antagonist drug in a solution (e.g., blood plasma or serum). For example, host cells may be transformed with a recombinant expression construct of the present invention, Y5 protein expressed in those host cells, the cells lysed, and the membranes from those cells used to screen compounds for Y5 binding activity. Competitive binding assays in which such procedures may be carried out are well known in the art. By selection of host cells which do not ordinarily express Y5, pure or crude preparations of

membranes containing Y5 can be obtained. Further, Y5 agonists and antagonists can be identified by transforming host cells with a recombinant expression construct as provided by the present invention. Membranes obtained from such cells (and membranes of intact cells) can be used in binding studies wherein the drug dissociation activity is monitored.

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It is known that the neurotransmitter NPY is a regulator of appetite. As shown herein, the various NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, bind with a high affinity to the Y5 receptor. Conversely, the NPY analogs/fragments that bind to the Y5 receptor with a lower affinity, such as (20-36)NPY, do not elicit feeding. It is therefore evident that by contacting the Y5 receptor with agonists and antagonists, feeding can be modulated. Accordingly, antagonists to the Y5 receptor, identified by the methods described herein, can be used to reduce appetite and hence treat obesity, diabetes and hyperlipidemia, and, conversely, agonists to the Y5 receptor can be used to treat conditions such as anorexia.

This invention provides a pharmaceutical composition comprising an effective amount of a agonist or antagonist drug identified by the method described herein and a pharmaceutically acceptable carrier. Such drugs and carrier can be administered by various routes, for example oral, subcutaneous, intramuscular, intravenous or intracerebral. The preferred route of administration would be oral at daily doses of about 0.01-100 mg/kg.

This invention provides a method of treating obesity, diabetes or hyperlipidemia, wherein the abnormality is improved by reducing the activity of Y5 receptor or blocking the binding of ligands to a Y5 receptor, which method comprises administering an effective amount of the antagonist-containing pharmaceutical composition described above to suppress the subject's appetite. Similarly, the invention also provides methods for treating diseases and conditions resulting from underfeeding and/or a loss of appetite, which method comprises administering an effective amount of an agonist-containing pharmaceutical composition described above to stimulate the subject's appetite.

The recombinant expression constructs of the present invention are useful in molecular biology to transform cells which do not ordinarily express Y5 to thereafter express this receptor. Such cells are useful as intermediates for making cell membrane preparations useful for receptor binding assays, which are in turn useful for drug screening. Drugs identified from such receptor assays can be used for the treatment of obesity, diabetes or anorexia.

The recombinant expression constructs of the present invention are also useful in gene therapy. Cloned genes of the present invention, or fragments thereof, may also be used in gene

therapy carried out by homologous recombination or site-directed mutagenesis. See generally Thomas & Capecchi, Cell 51, 503-512 (1987); Bertling, Bioscience Reports 7, 107-112 (1987); Smithies et al., Nature 317, 230-234 (1985).

Oligonucleotides of the present invention are useful as diagnostic tools for probing Y5 gene expression in tissues. For example, tissues are probed *in situ* with oligonucleotide probes carrying detectable groups by conventional autoradiographic techniques, as explained in greater detail in the Examples below, to investigate native expression of this receptor or pathological conditions relating thereto. Further, chromosomes can be probed to investigate the presence or absence of the Y5 gene, and potential pathological conditions related thereto, as also illustrated by the Examples below. Probes according to the invention should generally be at least about 15 nucleotides in length to prevent binding to random sequences, but, under the appropriate circumstances may be smaller.

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The invention also provides antibodies that are immunologically reactive to a mammalian Y5, preferably rat or human Y5. The antibodies provided by the invention are raised in animals by inoculation with cells that express a mammalian Y5 or epitopes thereof, using methods well known in the art. Animals that are used for such inoculations include individuals from species comprising cows, sheep, pigs, mice, rats, rabbits, hamsters, goats and primates. Preferred animals for inoculation are rodents (including mice, rats, hamsters) and rabbits. The most preferred animal is the mouse.

Cells that can be used for such inoculations, or for any of the other means used in the invention, include any cell line which naturally expresses a mammalian Y5, or any cell or cell line that expresses a mammalian Y5 or any epitope thereof as a result of molecular or genetic engineering, or that has been treated to increase the expression of a mammalian Y5 by physical, biochemical or genetic means. Preferred cells are human cells, most preferably HEK 293 and BHK cells that have been transformed with a recombinant expression construct comprising a nucleic acid encoding a mammalian Y5, preferably a rat or human Y5, and that express the mammalian Y5 gene product.

The present invention provides monoclonal antibodies that are immunologically reactive with an epitope of mammalian Y5 or fragment thereof and that is present on the surface of mammalian cells, preferably human or mouse cells. These antibodies are made using methods and techniques well known to those of skill in the art.

Monoclonal antibodies provided by the present invention are produced by hybridoma cell lines, that are also provided by the invention and that are made by methods well known in the art. Hybridoma cell lines are made by fusing individual cells of a myeloma cell line with spleen cells derived from animals immunized with cells expressing the Y5 receptor, preferably rat or human cells, as described above. The myeloma cell lines used in the invention include lines derived from myelomas of mice, rats, hamsters, primates and humans. Preferred myeloma cell lines are from mouse. The animals from whom spleens are obtained after immunization are rats, mice and hamsters, preferably mice, most preferably Balb/c mice. Spleen cells and myeloma cells are fused using a number of methods well known in the art, including but not limited to incubation with inactivated Sendai virus and incubation in the presence of polyethylene glycol (PEG). The most preferred method for cell fusion is incubation in the presence of a solution of 45% (w/v) PEG-1450. Monoclonal antibodies produced by hybridoma cell lines can be harvested from cell culture supernatant fluids from in vitro cell growth; alternatively, hybridoma cells can be injected subcutaneously and/or into the peritoneal cavity of an animal, most preferably a mouse, and the monoclonal antibodies obtained from blood and/or ascites fluid.

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Monoclonal antibodies provided by the present invention are also produced by recombinant genetic methods well known to those of skill in the art, and the present invention encompasses antibodies made by such methods that are immunologically reactive with an epitope of a mammalian Y5.

The present invention encompasses fragments of the antibody that are immunologically reactive with an epitope of a mammalian Y5. Such fragments are produced by any number of methods, including but not limited to proteolytic cleavage, chemical synthesis or preparation of such fragments by means of genetic engineering technology. The present invention also encompasses single-chain antibodies that are immunologically reactive with an epitope of a mammalian Y5 made by methods known to those of skill in the art.

The present invention also encompasses an epitope of a mammalian Y5 that is comprised of sequences and/or a conformation of sequences present in the mammalian Y5 molecule. This epitope may be naturally occurring, or may be the result of proteolytic cleavage of the mammalian Y5 molecule and isolation of an epitope-containing peptide or may be obtained by synthesis of an epitope-containing peptide using methods well known to those skilled in the art. The present invention also encompasses epitope peptides produced as a result

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of genetic engineering technology and synthesized by genetically engineered prokaryotic or eukaryotic cells.

The invention also includes chimeric antibodies, comprised of light chain and heavy chain peptides immunologically reactive to an epitope that is a mammalian Y5. The chimeric antibodies embodied in the present invention include those that are derived from naturally occurring antibodies as well as chimeric antibodies made by means of genetic engineering technology well known to those of skill in the art.

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Also provided by the present invention are non-human transgenic animals grown from germ cells transformed with the Y5 nucleic acid sequence according to the invention and that express the Y5 receptor according to the invention and offspring and descendants thereof. Also provided are transgenic non-human mammals comprising a homologous recombination knockout of the native Y5 receptor, as well as transgenic non-human mammals grown from germ cells transformed with nucleic acid antisense to the Y5 nucleic acid of the invention and offspring and descendants thereof. Further included as part of the present invention are transgenic animals which the native Y5 receptor has been replaced with the human homolog. Of course, offspring and descendants of all of the foregoing transgenic animals are also encompassed by the invention.

Transgenic animals according to the invention can be made using well known techniques with the nucleic acids disclosed herein. E.g., Leder et al., U.S. Patent Nos.4,736,866 and 5,175,383; Hogan et al., Manipulating the Mouse Embryo, A Laboratory Manual (Cold Spring Harbor Laboratory (1986)); Capecchi, Science 244, 1288 (1989); Zimmer and Gruss, Nature 338, 150 (1989); Kuhn et al., Science 269, 1427 (1995); Katsuki et al., Science 241, 593 (1988); Hasty et al., Nature 350, 243 (1991); Stacey et al., Mol. Cell Biol. 14, 1009 (1994); Hanks et al., Science 269, 679 (1995); and Marx, Science 269, 636 (1995). Such transgenic animals are useful for screening for and determining the physiological effects of Y5 receptor agonists and antagonist. Consequently, such transgenic animals are useful for developing drugs to regulate physiological activities in which NPY and/or PYY participate.

The following Examples are provided for illustrative purposes only and are not intended, nor should they be construed, as limiting the invention in any manner.

EXAMPLES

Example 1

Isolation and Sequencing of Rat Y5 Receptor

Isolation of rat hypothalamus mRNA and construction of cDNA library

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Expression cloning strategy was used to clone novel NPY receptor in rat hypothalamus cDNA library. RNA was obtained from 9 frozen rat hypothalami weighing a total of 0.87 grams. Poly(A) RNA was isolated directly from the tissue using the Promega PolyATtract System 1000 kit (Promega, Madison, WI). The hypothalami were homogenized in 4 mL of 4M guanidine thiocyanate-25mM sodium citrate, pH 7.1-2% B-mercaptoethanol using a Polytron at full-speed for approximately 1 minute. To the homogenized tissue 8 mL of 4M guanidine thiocyanate-25mM sodium citrate, pH 7.1-1% \(\beta\)-mercaptoethanol which had been preheated to 70°C was added. After mixing thoroughly, 870 pmol biotinylated oligo(dT) was added; the mixture was incubated at 70°C for 5 minutes. The homogenate was subjected to centrifugation at 12000 x g for 10 minutes at room temperature; the homogenate was transferred to a clean tube and 10.44 mL Streptavidin MAGNESPHERE® Paramagnetic Particles (SA-PMPs) which had been prepared as per the published protocol was added. (Promega Corp. published protocol TM 228; Promega Corporation, Madison, WI). The homogenate and SA-PMPs were incubated together for 2 minutes at room temperature after which the homogenate was decanted while the SA-PMP-biotinylated oligo(dT)-hypothalamic poly(A) RNA complex was retained in the tube by a magnetic stand. The complex was washed as per the protocol, after which the RNA was precipitated and resuspended in water. 25 micrograms of this poly(A) RNA was used by Invitrogen (Invitrogen Corporation, San Diego, CA) to prepare a cDNA expression library. The protocols used by Invitrogen to prepare the cDNA library are essentially based upon the procedures of Okayama and Berg (Molec. Cell. Biol. 2, 161 (1982)) and Gubler and Hoffman (Gene 25, 263 (1983)) (Invitrogen Corporation publications 130813sa and 130928sa). An oligo(dT) anchor primer was used for reverse transcription, and the library was cloned unidirectionally into pcDNA3 vector which contains a CMV promoter for eukaryotic expression. The cDNA library had 5.3 x 10⁵ primary recombinants with an average insert size of 2.59 kb.

Isolation of a novel Y5 receptor cDNA clone

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The rat hypothalamus cDNA library was plated on the LB/Ampicillin plates in pools of 1,000 independent colonies. The plates were incubated at 37°C for about 20 hours and the bacteria from each plate were scraped in 4-5 ml LB/Ampicillin media. Two ml of the bacteria samples were used for plasmid preparation and one ml of each pool was stored at -80°C in 15% glycerol.

COS-7 cells were grown in Dulbecco's Modified Eagle Medium (DMEM, GIBCO 11965-092), 10% fetal bovine serum (GIBCO 16000-028), and 1 x antibiotic/antimycotic solution (GIBCO 15240-039) (Gaithersburg, MD). Cells were trypsinized and split at 50 to 70% confluency.

DNA from 1300 pools was transfected into COS-7 cells for [125I]PYY binding assays. Twenty four hours before transfection, cells were plated into flaskette chambers (Nunc, Inc. 177453, Naperville, IL) at 3x10⁵ cells/flaskette (equivalent to 3x10⁴ cells/cm²). Two µg of plasmid DNA from each pool was transfected into the cells using 10 µl of Lipofectamine (GIBCO 18324-012) according to the manufacture's protocol. Forty eight hours after transfection, the [125I]PYY binding assay was performed in the flaskette chamber. The cells were treated with 2 ml total binding buffer: 10 mM HEPES, 5 mM KCl, 1.2 mM KH₂PO₄, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 150 mM NaCl, 25 mM NaHCO₃, 10 mg/ml bovine serum alburnin, 0.5 mg/ml bacitracin and 0.4 mg/ml soybean trypsin inhibitor at room temperature for 15 minutes. The cells were then incubated with 100 pM porcine [125I]PYY (Amersham (Arlington Heights, IL), Specific Activity 4000Ci/mmol) in the total binding buffer for 90 minutes at room temperature. After binding, the cells were washed three times with ice-cold total binding buffer without ligand and one time with cold phosphate buffered saline (PBS). Cells were fixed with 1% cold glutaraldehyde in PBS for 15 minutes, washed once with cold PBS/0.5 M Tris, pH 7.5 and incubated in PBS/0.5 M Tris, pH 7.5 for 15 minutes at 4°C. After washing one more time with cold PBS, the slides were dipped in 0.5% gelatin at 42°C and dried The dried slides were dipped in 50% photographic emulsion (Kodak (Rochester, NY) NTB2) at 42°C and exposed in the darkbox for four days at 4°C. After four days of exposure, the darkbox was moved to room temperature for one hour and slides were developed in developer D-19 (Kodak) for three minutes at 15°C and fixed in fixer (Kodak) for three minutes at 15°C, washed in water and air dried. Cells were stained with Diff-Quik stain

set (Baxter, McGaw Park, IL) and air dried. Slides were dipped into xylenes and mounted with DPX mountant (Electron Microscopy Science, Fort Washington, PA). Positive cells were identified using dark field microscopy.

Twenty one positive pools were identified. Since the hypothalamus expresses different subtypes of NPY receptors including Y1 and Y2 receptors, we analyzed all the positive pools for Y1, Y2 and Y4/PP receptors by PCR. Of the 21 positive pools tested as described above, 12 pools contained Y1, 4 pools contained Y2 and none contained Y4/PP. Five pools (Y217, Y555, Y589, Y861 and Y1139) were negative by PCR analysis. The pool Y217 was subdivided in 24 subpools of 200 colonies, then 50 colonies, and finally a single clone, the Y217.24.13.6B clone (6B), was isolated.

DNA and peptide sequences analysis

Plasmid DNA was sequenced by Lark Technologies Inc. (Houston, Texas) and Biotechnology Resource Laboratory of Yale University (New Haven, CT) using Sequenase Kit (US Biochemical, Cleveland, OH) or Applied Biosystems' automatic sequencer system (model 373A). The peptide sequence was deduced from the long open-reading-frame of the nucleotide sequence. DNA and peptide sequences were analyzed using the GCG program (Genetics Computer Group, Madison, WI). The results are embodied in SEQ ID NO 1 (the nucleic acid sequence) and SEQ ID NO 2 (the amino acid sequence).

Example 2

Localization of Rat Y5 Receptor in Brain and Other Tissues

Northern Blot

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To study the expression level of the Y5 receptor in the rat brain and other tissues, we did Northern blot analysis using the 6B 2.4 kb probe. A rat multiple tissue Northern blot (Clontech Laboratories, Palo Alto, CA) was hybridized to the ³²P-labeled rat 6B probe. The blot contains 2 μg of poly A⁺ RNA per lane from rat heart, brain, spleen, lung, liver, skeletal muscle, kidney, and testis. Hybridization was carried out in 1x hybridization solution containing 6X SSC (0.9 M NaCl, 0.09 M Na Citrate, pH 7.0), 5x Denhardt's solution (0.1% polyvinylpyrrolidone, 0.1% ficoll type 400, 0.1% bovine serum albumin), 100 mg/ml sheared, and denatured salmon sperm DNA at 65°C. The filter was washed at 65°C in 0.1X SSC, 0.1% SDS and exposed to Kodak XAR 5 film with two intensifying screens. A single 2.6 kb band was detected in the brain after

overnight exposure of the blot. No bands were found from other tissues (heart, spleen, lung, liver, skeletal muscle, kidney and testis) in the Clontech multiple tissue Northern blot, even after six days of exposure.

We tested 6B expression in more rat tissues and different regions of brain. mRNA was isolated from rat whole brain, cortex, hypothalamus, hippocampus, olfactory bulb, spleen, stomach, kidney, small intestine, adrenal and pancreas using Fast Track Isolation Kit (Invitrogen). Ten µg of mRNA from different brain regions and multiple tissues were run on a denaturing formaldehyde 1% agarose gel, transferred to a Nytran membrane (Schleicher and Schuell) and hybridized with ³²P-labeled 6B 2.4 kb probe and washed at high stringency. After overnight hybridization, the filter was washed at high stringency and exposed to X-ray film with intensifying screens. The 6B receptor mRNA was detectable in the brain regions examined after one day exposure, but no signal was observed from other tissues, even after a week exposure with double intensifying screens.

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Example 3

Isolation of Two Isoforms of the Rat Y5 Receptor

Plasmid DNA from pools Y555, Y589, and Y861 described in Example 1 were hybridized to the Y5 probe at high stringency. A single positive clone was isolated from the Y555 pool and sequenced as described in Example 1. Compared to the 6B DNA sequence, the Y555 sequence (SEQ ID NO 4 has a 123 bp insert sequence located at the 5'-untranslated region between nucleotides 239 and 240 of Y5 clone. The coding region of the clones Y555, Y589, and Y861 has the same sequence as clone 6B, except for one nucleotide substitution (C to T) at position 430 of the 6B clone. The nucleotide substitution changes the amino acid proline to leucine in the first transmembrane domain. The corresponding amino acid sequence is given by SEQ ID NO 4.

The different isoforms of the receptor could be the allelic variants of the same gene. To test this hypothesis, we analyzed genomic DNA from 16 rats. The genomic DNA from each animal was used as template for PCR analysis. A 314 bp DNA fragment that contains the site of the nucleotide variation was amplified and sequenced. Of the 16 DNA samples tested, 14 samples had a T at position 430 and 2 samples had a C. This result strongly suggests that the amino acid variation is an allelic variant.

Example 4

Pharmacological Characterization of the Novel Rat NPY Receptors

Transient Transfection

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Monkey kidney cells (COS-7) were maintained in T-175 cm² flasks (NUNC) at 37°C with 5% CO₂ in a humidified atmosphere. Cells were grown in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 2 mM glutamine, 10% fetal bovine serum, 1 mM sodium pyruvate, and antibiotic/antimycotic. Cells at 70% confluency were transfected with Y5 DNA using the Lipofectamine method (GIBCO-BRL). 15 μg DNA and 90 μl Lipofectamine were added to each flask. Media was completely replaced 24 hours post transfection, and membranes were harvested 24 hours later.

Stable Expression of the Rat NPY Y5 receptor (clone Y861)

A strain of the human embryonic kidney cell line 293 adapted to grow in suspension (293S) was used for these experiments. Approximately 1x10⁶ cells were seeded onto a 10-cm dish 24 hours prior to transfection. The rat NPY Y5 cDNA (Y861), subcloned in the eukaryotic expression vector pcDNA3 (Invitrogen, Carlsbad, CA) was first linearized with Notl and purified using a Wizard PCR Prep kit (Promega). In preparation for transfection, 15 µg of the linearized DNA were added to 500 µl of DMEM cell culture media, and 30 µl of Lipofectamine (Life Sciences) were added into a separate 500 µl aliquot of DMEM. These two solutions were mixed together and incubated for 20 minutes at room temperature and the resulting DNA/lipid complexes were then slowly added to the cells (which had been previously rinsed once with serum-free DMEM) and covered with a total volume of 10 ml. Cells were then transferred to a humidified 10% CO₂ incubator and left for 4 hours at 37°C, at which time the media was replaced with DMEM supplemented with 8% FBS. After 16 hours, cells were trypsinized and split at a 1:15 ratio into 10-cm dishes containing DMEM/8% FBS in the presence of 700 μg/ml of G418 (selection media). When discrete colonies became apparent (after approximately 10 days), cells were pooled and carried through 2 additional passages in selection media. Cells were then trypsinized and diluted in preparation for cloning by limited dilution (CBLD), such that an average of one cell was seeded in each well of a 96-well microtiter culture plate, and was inspected periodically for the subsequent 2 to 3 weeks. After 21 days in culture under selection conditions, those wells containing single colonies were selected and transferred to 24well culture plates following trypsinization. Each of these clones was propagated until

sufficient quantities were available for testing [125]PYY binding activity, from which one particular clone designated E7 was selected on the basis of its high level of binding activity.

Stable Expression of the human NPY Y5 receptor

293 cells were plated onto a T75 flask one day prior to transfection such that they were 50-70% confluent when used for the experiment. The human NPY Y5 intronless genomic clone HG.PCR15, containing the full length open reading frame encoding the receptor, was first linearized with *Not* I and purified using a Wizard PCR Prep kit (Promega). For each transfection, 8 μg of linearized DNA were added to 1.25 ml of Optimem culture media (Life Sciences) and 37 μl of Transfectam (Promega) were added to 1.25 ml of Optimem. These two solutions were then mixed together and added to cells previously washed once with Optimem. After an incubation period of 5 hours, the DNA/Transfectam mixture was removed, cells were washed with PBS and fed with DMEM supplemented with 10% FBS. Cells were left intact for two days, and then switched to selection media (DMEM 10% FBS containing 350 μg/ml of G418) for 5-10 days followed by CBLD as described above. The individual clone 293.hy5.sb.8 was selected on the basis of its high level of [125]PYY binding activity, using the intact cell binding protocol from above.

Membrane Preparation

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The media was removed from each flask of transfected cells, and the cells were washed twice with 20 ml ice-cold phosphate buffered saline. The cells were scraped from the flask in 5 ml of Tris buffer (20 mM Tris-HCl and 5 mM EDTA, pH 7.7), and then transferred to a centrifuge tube. Each flask was washed with an additional 5 ml of Tris buffer and combined in the centrifuge tube. The cells were polytroned for 2 x 10 seconds (12 mm probe, 7000-8000 rpm) and centrifuged 5 minutes (Centra 7R, International Equipment Co., Needham Heights, MA) at 800 rpm and 4°C. The supernatant was then transferred to a clean centrifuge tube and was centrifuged at 30,000 x g for 30 minutes and 4°C. The supernatant was removed and the pellet was stored at -80°C. Protein concentration was measured using the Bio-Rad kit pursuant to the standard manufacturer's protocol (Biorad Laboratories, Hercules, CA) with bovine IgG as the standard.

[125] [11PYY Binding Assay for NPY Y5 Receptors

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The binding assays were performed on GF/C Millipore (Bedford, MA) 96-well plates pretreated with 0.02% polyethylenimine (PEI) for at least 2 hours prior to use. The PEI was aspirated from the plates on a vacuum manifold immediately before the samples were added to the wells. All peptides, tissue and radioligand were diluted with binding buffer (25 mM Tris, 120 mM NaCl, 5 mM KCl, 1.2 mM KH₂PO₄, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 0.1% BSA and 0.5 mg/ml bacitracin, pH 7.4). For competition assays, increasing concentrations of peptide were incubated with [¹²⁵I]PYY and tissue. In a final volume of 200 μl, samples consisted of: membrane protein (*i.e.*, 2.5-15 or 10-30 μg membrane protein for rat Y5 or human Y5, respectively); 75-100 pM [¹²⁵I]PYY NEN-DuPont (Boston, MA); peptide dilution or binding buffer. Nonspecific binding was defined by 1 μM PYY. NPY, PYY, (2-36)NPY, (10-36)NPY, (LP)(3-36)NPY and (32D-Trp)NPY were synthesized at Bayer Corp. (West Haven, CT). All other peptides were purchased from either Peninsula (Belmont, CA) or Bachem (Torrance, CA).

For saturation experiments, increasing concentrations of [¹²⁵I]PYY were incubated with membrane and 1 µM PYY. After a 2 hour incubation at room temperature with constant mixing, the samples were aspirated on a vacuum manifold. The wells were washed with three 200 µl aliquots of ice-cold binding buffer. The individual wells were punched into 12x75 mm plastic tubes, and counted on a Wallac (Gaithersburg, MD) gamma counter. Binding data were analyzed using the nonlinear regression curve-fitting program RS/1 (BBN Software Products Corp., Cambridge, MA).

Binding Assays for Rat Y2. Y1. and Y4/PP1 Receptors

The binding buffer for rat Y2 binding was Krebs/Ringer bicarbonate (Sigma K-4002, S-8875), pH 7.4, containing 0.01% bovine serum albumin (BSA - Sigma A-2153) and 0.005% bacitracin. 0.85-1 μ g of protein and 25 pM [125 I]PYY are added to each well. Nonspecific binding is defined by 1 μ M NPY.

The binding buffer for rat Y1 and rat Y4/PP1 binding consisted of 137 mM NaCl, 5.4 mM KCl, 0.44 mM KH₂PO₄, 1.26 mM CaCl₂, 0.81 mM MgSO₄, 20 mM HEPES, 1 mM dithiothreitol (DTT), 0.1% bacitracin, 100 mg/l streptomycin sulfate, 1 mg/l aprotinin, 10 mg/ml soybean trypsin inhibitor and 0.3% BSA, pH 7.4. For rat Y1 binding, ~5-15 μg of protein and 50 pM [¹²⁵I]PYY were added to each well, and nonspecific binding was defined by 1 μM NPY. For the rat Y4/PP1 binding assay, ~1-2 μg of protein and 50 pM rat [¹²⁵I]PP (NEN

DuPont, Boston, MA) were added to each well, and 1 μ M rat PP was used to define nonspecific binding.

In Vitro Functional Assay - Measurement of Forskolin-Stimulated Adenylate Cyclase

Rat Y5

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(Reference: Gordon et al., J. Neurochem. 55, 506, 1990) Suspension cells stably expressing the Y5 receptor (approximately 400,000 per sample) were resuspended in serum-free DMEM containing 10mM HEPES (pH 7.4) and 1 mM isobutylmethylxanthine (IBMX). 1uM forskolin was added to the cells. The assay was stopped by transferring the samples into a boiling water bath for 3 minutes. After a 3 minute centrifugation at 14,000xg, an aliquot of each sample was quantitated for cAMP levels by radioimmunoassay (NEN DuPont, MA).

Human Y5

Monolayer cells stably expressing the Y5 receptor were pre-rinsed with Wash buffer (pH 7.2: 137 mM NaCl, 2.7 mM KCl, 0.9 mM CaCl₂, 0.5 mM MgCl₂, 6.5 mM Na₂HPO₄, 1.5 mM KH₂PO₄). Cells were then incubated for 10 minutes at 37°C in Assay buffer (pH 7.4: Wash buffer + 10 mM HEPES, 10 mg/ml BSA, 0.5 mg/ml bacitracin, 0.4 mg/ml soybean trypsin inhibitor). After addition of fresh buffer and 100 μM IBMX, the cells were incubated for 10 minutes at 37°C. The reaction was started with the addition of peptide and 1-10 μM forskolin. After a 20 minute incubation at 37°C, the reaction was terminated by discarding the buffer and adding 65% ethanol to each well. The supernatant was then transferred to microfuge tubes and the extraction step was repeated once more. After evaporation of the ethanol from the samples, the amount of cAMP was assayed using by radioimmunoassay (NEN DuPont: Boston, MA).

In Vivo Pharmacology Procedures

Adult male Wistar rats were surgically implanted with a chronic intracerebral ventricular (ICV) cannula (Plastic Products, Roanoke, Virginia) using a stereotaxic instrument. Several days after the surgery, 1-6 nmoles of each peptide (or saline) was injected into the lateral ventricle of 4-12 rats in a volume of 5-10 μl. The quantity of rodent chow consumed in a 2 hour period was measured.

In Vitro and In Vivo Pharmacology Results

Figure 1 presents the competition curves of various peptides for [125 I]PYY binding to Y5 receptor membranes transiently expressed in COS-7 cells. Each point is the average value of triplicate determinations from a representative experiment. IC₅₀ values corresponding to 50% inhibition of specific binding were determined using nonlinear regression analysis. K_i values were calculated from the IC₅₀ values using the Cheng-Prusoff correction, such that $K_i = IC_{50}/(1 \pm (L/K_d))$, where L is the radioligand concentration and K_d is the dissociation constant. The results for transiently expressed Y5 clones are presented in Table 1, and Table 2 contains data for stably expressed Y5 clones.

Table 1

PEPTIDE	NPY Y	5 BINDING AFF	INITIES (K _i ± S	SEM, nM)
	RAT Y555 Clone*	RAT 6B Clone	RAT Y861 Clone	HUMAN Clone
r/hNPY	0.53 ± 0.06	0.49 ± 0.03	0.50 ± 0.06	0.73 ± 0.09
rPYY	1.1 (1.2, 0.95)	0.48 ± 0.09	1.0 ± 0.13	1.3 ± 0.14
h(LP)PYY	2.5 ± 0.5	0.57 ± 0.01	1.8 ± 0.09	1.7 ± 0.3
r/h(LP)NPY	0.96 (1.0, 0.92)	0.31	0.55 ± 0.11	0.97 ± 0.36
p(LP)NPY	ND	0.64 ± 0.07	0.47	0.88 ± 0.11
r/h(2-36)NPY	0.81 (0.61, 1)	0.65	1.2 ± 0.07	1.2 ± 0.15
p(3-36)NPY	3.6 ± 0.4	1.9 ± 0.27	2.0 (1.8, 2.3)	10.4 ± 2.0
r/h(3-36)NPY	ND	0.49	2.1 (2.7, 1.6)	3.8 ± 0.48
r(3-36)PYY	6.2 ± 1.1	1.4 ± 0.10	4.2 ± 0.47	10 ± 3.4
r/h(10-36)NPY	35	4.9 (6.0, 3.8)	34 ± 2.8	110 (110, 109)
p(13-36)NPY	40 (38, 41)	7.7 (7.9, 7.5)	22 (25, 19)	56 <u>+</u> 7
r(13-36)NPY	73	11 ± 1.0	86 ± 19	77 (89, 65)
p(18-36)NPY	303	194 ± 88	206 ± 61	618 ± 85
r/h(20-36)NPY	636	330 ± 31	587	>1000
r/h(22-36)NPY	>1000	>1000	>1000	>1000
r/h(26-36)NPY	>1000	>1000	>1000	>1000
(1-24)NPY	ND	>1000	>1000	>1000
BIBP3226	ND	>1000	>1000	>1000
hPP	ND	ND	4.0 ± 0.29	11 (15, 6.2)
гРР	ND	62	296 ± 47	436 (582, 290)

^{*} IC₅₀ values (nM)

Table 2

Peptide	K; Values (nM; A	verage ± SEM)
	293.hY5.sb.8	293S.Y861.2
rPYY	1.3 ± 0.2	0.71 ± 0.1
hPYY	1.1 ± 0.2	1.06 ± 0.2
(3-36)PYY	4.5 ± 0.7	3.6 ± 0.4
(13-36)PYY	24 ± 2.0	29 ± 4
h(LP)PYY	1.3 ± 0.1	0.76 ± 0.1
r/hNPY	0.79 ± 0.1	0.86 ± 0.07
p(LP)NPY	1.2 ± 0.4	0.67 ± 0.04
h/r(LP)NPY	0.89 ± 0.1	0.67 ± 0.04
(LP)(3-36)NPY	3.1 ± 0.6	2.9 ± 0.9
(2-36)NPY	1.4 ± 0.03	0.83 ± 0.1
h(3-36)NPY	3.5 ± 0.4	1.4 ± 0.4
(10-36)NPY	14 ± 2.7	15 ± 4.7
p(13-36)NPY	8.7 ± 1.6	8.8 ± 2.0
p(18-36)NPY	144 ± 18	61 ± 13
(20-36)NPY	429 ± 133	108 ± 16
(22-36)NPY	>900	>930
(26-36)NPY	>900	>930
(1-24)NPY	>900	>930
(32D-Trp)NPY	7.3 ± 0.8	4.2 ± 1.0
hPP	3.7 ± 1.6	2.5 ± 0.5
rPP	286 ± 77	203 ± 44

 K_1 values for various peptides for [125 I]PYY binding to the transiently expressed rat 6B, Y861 and Y555 receptor clones as well as the human Y5 receptor. The averages \pm standard error of the mean (SEM) represent values from at least three independent experiments. Two independent experiments are represented by the average, followed by the individual values in parentheses. Remaining values without SEM are from a single experiment. Peptide species in Table 1 (and Table 2, *infra*) are indicated with the following prefixes: r = rat, h = human, p = porcine, r/h = rat = human. ND = not determined.

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The rank order of the affinities of the peptides tested is as follows:

In Table 3, the pharmacological profile of the standard peptides is expanded for the other cloned NPY receptors to further illustrate the novel nature of the Y5 receptor pharmacology. In addition, the *in vivo* feeding response of some of these peptides is listed for comparison. The data shown are representative of the average of at least two independent experiments, as described in the methods. Feeding of rats injected (ICV) with saline was < 3g/2hours.

Table 4 shows the EC_{50} values for same standard peptides at the rat and human Y5 receptor.

C-terminal fragment (3-36)NPY binds preferentially to Y2 receptors, while (LP)NPY has lower affinity. Conversely, (LP)NPY has high affinity for the Y1 receptor, while (3-36)NPY and the C-terminal fragments are much weaker. When considering the rat Y4/PP1 receptor, rat PP has very high affinity as compared to NPY, PYY, (LP)NPY, and (13-36)NPY. In the *in vivo* feeding model, (LP)NPY, which has high affinity for Y1 and low affinity for Y2, and (3-36)NPY, which has a high affinity for Y2, but not Y1, all stimulate feeding in rats. Rat PP does not induce much feeding when administered to rats. This *in vivo* profile matches the *in vitro* pharmacological profile outlined in Table 2 for the Y5 receptor.

In addition, while (LP)(3-36)NPY (a custom peptide synthesized at Bayer) has weak affinity for Y1, Y2 and Y4/PP1, it stimulates feeding in rats. Importantly, (LP)(3-36)NPY has high affinity for the Y5 receptor (Table 2). These data are further evidence that the Y5 receptor is linked to feeding.

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Table 3

IC ₅₀ VALUE (nM)												
PEPTIDE	Rat Y1 (clone)	Rat Y2 (clone)	Rat Y4/PP1 (clone)	Rat Y5 (Y861)	Feeding (g/2 h)							
r/hNPY	0.13	0.24	> 1000	0.45	>5							
rPYY	0.43	0.079	630	0.9	>5							
n(Leu ³¹ Pro ³⁴)PYY	0.57	116	ND	2.0	>5							

IC ₅₀ VALUE (nM)												
PEPTIDE	Rat Y1 (clone)	Rat Y2 (clone)	Rat Y4/PP1 (clone)	Rat Y5 (Y861)	Feeding (g/2 h)							
p(Leu ³¹ Pro ³⁴⁾ NPY	0.15	150	4.3	0.63	>5							
r/h(2-36)NPY	47	0.50	>1000	1.3	>5							
p(3-36)NPY	45	0.67	>1000	2.2	>5							
r/h(Leu ³¹ Pro ³⁴) (3-36)NPY	44	154	20	3.4	>5							
hPP	40	>1000	0.065	4.9	>5							
(32DTrp)NPY	> 1000	26	ND	7.0	ND							
r/h(10-36)NPY	148	0.42	>1000	34	<3							
rPP	843	>1000	0.071	325	<3							
p(18-36)NPY	287	0.34	159	326	<3							
(20-36)NPY	435	0.64	ND	638	<3							
(22-36)NPY	>1000	0.89	ND	>1000	<3							
(26-36)NPY	>1000	84	ND	>1000	<3							
(1-24)NPY	>1000	>1000	ND	>1000	<3							

The pharmacological profile for the 6B (and Y861 and Y555) receptor clones is distinct from Y1 receptors (where PYY~NPY~(LP)NPY > (3-36)NPY > (13-36)NPY ~ (18-36)NPY > (LP)(3-36)NPY), as well as Y2 receptors (where PYY~NPY~(13-36)NPY~(18-36)NPY~(3-36)NPY >> (LP)NPY~(LP)(3-36)NPY). The Y5 receptor is also different from the pancreatic polypeptide (PP) receptor (Y4/PP) since [1251] PP (rat) does not bind to it.

Although the rank order of affinities is essentially the same when comparing 6B to Y861 and Y555, subtle differences do exist in the IC_{50} values. It appears that Y861 and Y555 have slightly lower affinities (approximately 2- to 3-fold) for PYY and other PYY analogs, as compared to 6B. In addition, (10-36)NPY and (13-36) have 2- to 4-fold lower affinity for Y861 and Y555.

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Nonlinear regression analysis of saturation data for the Y5 receptor yielded a K_d value of 0.27 nM and a receptor density (B_{max}) of about 140 fmol/mg protein in these transiently transfected cells.

Fig. 2 presents the saturation curve for specific binding of [¹²⁵I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells. Membranes were incubated with concentrations of [¹²⁵I]PYY ranging from 0.05 to 5 nM, in the presence or absence of 1 μM PYY. Each point represents the average value of triplicate determinations at each concentration tested. The inset in Fig. 2 shows the corresponding Rosenthal plot of the data.

Table 4

Peptide	EC ₅₀ Value	s, nM (n Value)
	293S.Y861.2	293.hY5.sb.8
r/hNPY	6.3 ± 1.9 (3)	0.3 (1)
rPYY	6.5 (2)	ND
r/h(2-36)NPY	21 (2)	ND
r/h(3-36)NPY	ND	6 (1)
r/h(LeuPro)(3-36)NPY	31 ± 39 (3)	23 ± 11 (3)
(32D-Trp)NPY	24 (1)	33 (1)
hPP	1 (1)	5 (1)
rPP	112 (1)	>1000 (1)

Example 5 Isolation of Human Y5 Receptor

10 Isolation of Human Genomic Clone

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Polymerase chain reaction (PCR) was used to amplify a 375 base pair (bp) coding region of the rat Y5 cDNA clone. The primers for the PCR were:

- (+) 5'-TAGGGAACCTGGCCTCCTCC-3' (SEQ ID NO 5) (nucleotides 487-506),
- (-) 5'-TCAGAGGGCCATGACTCAAC-3' (SEQ ID NO 6) (nucleotides 843-862).
- The PCR product was cloned into pCRII vector (Invitrogen) and sequenced. After confirmation by sequencing, the insert was purified from the low melting gel and labeled with digoxigenin-l1-dUTP using the random primed method (Boehringer Mannheim, Indianapolis, IN). The labeled probe was used to screen human genomic library.

1x10⁶ independent recombinants were screened from the library. Filter hybridization was carried out in the hybridization buffer containing 6x SSC, 0.1% N-lauroylsarcosine, 0.02% sodium dodecyl sulfate (SDS), 3% blocking reagent (Boehringer Mannheim) and 30% formamide at 37°C overnight. The filters were washed at 37°C in 0.1x SSC, 0.1% SDS and the positive clones were identified by CSPD detection kit according to the manufacturer's protocol (Boehringer Mannheim).

Two positive clones (HG11A and HG19) were isolated from the library. The positive clones were subcloned into pBluescript vector (Stratagene). One clone, h11a, was analyzed by restriction mapping and plasmid Southern blot. Two EcoRV fragments, 2.4 kb and 0.4 kb, were hybridized by the rat Y5 probe. These two DNA fragments were subcloned and sequenced from both ends. DNA sequence analysis was performed using GCG program. The coding region of the human Y5 genomic clone was identified by DNA sequence analysis. This region was amplified by PCR using genomic clone h11A as template and subcloned into pcDNA3 expression vector (Invitrogen) for further studies. The h11A clone has the nucleic acid coding sequence given by SEQ ID NO 5 and the protein that it encodes has the amino acid sequence given by SEQ ID NO 6.

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The human Y5 DNA coding region was used to search the sequence similarities in the gene bank. The Y5 coding sequence from nucleotide 821 to the stop codon at position 1338 is nearly identical, but in an opposite orientation, to part of the human NPY-Y1 gene (Ball et al, J. Biol. Chem. 270, 30102 (1995)). The identical sequence covered the 1C exon promoter, exon 1C, and part of the intron sequences of the NPY-Y1 receptor in an opposite orientation. Compared to the published nucleotide sequence, the Y5 coding region has a T insertion at position 1226 and a TG insertion at positions 1235 and 1236.

SEQUENCE LISTING

	(1) GENE	ERAL INFORMATION:
5	(3)	ADDITION IN THE PARTY William
	(1)	APPLICANT: Hu Ph.D., Yinghe McCaleb Ph.D., Michael L.
		Bloomquist Ph.D., Brian T.
		Flores-Riveros Ph.D., Jaime R.
10		Cornfield Ph.D., Linda J.
	(ii)	TITLE OF INVENTION: Neuropeptide Y Receptor and Nucleic Acid Sequences
15	(iii)	NUMBER OF SEQUENCES: 8
	(iv)	CORRESPONDENCE ADDRESS:
		(A) ADDRESSEE: McDonnell Boehnen Hulbert & Berghoff
		(B) STREET: 300 South Wacker Drive
20		(C) CITY: Chicago
		(D) STATE: IL
		(E) COUNTRY: USA
		(F) ZIP: 60606
25	(v)	COMPUTER READABLE FORM:
		(A) MEDIUM TYPE: Floppy disk
		(B) COMPUTER: IBM PC compatible
		(C) OPERATING SYSTEM: PC-DOS/MS-DOS
		(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
30		
	(vi)	CURRENT APPLICATION DATA:
		(A) APPLICATION NUMBER:
		(B) FILING DATE: (C) CLASSIFICATION:
35		(C) CLASSIFICATION:
	(viii)	ATTORNEY/AGENT INFORMATION:
		(A) NAME: Greenfield Ph.D., Michael S.
		(B) REGISTRATION NUMBER: 37,147
		(C) REFERENCE/DOCKET NUMBER: 96,149/WH 405
40		
	(ix)	TELECOMMUNICATION INFORMATION:
		(A) TELEPHONE: (312)715-1000
		(B) TELEFAX: (312)715-1234
45		
	(2) INFO	RMATION FOR SEQ ID NO:1:
	(i)	SEQUENCE CHARACTERISTICS:
**		(A) LENGTH: 2481 base pairs
50		(B) TYPE: nucleic acid
		(C) STRANDEDNESS: single
		(D) TOPOLOGY: linear
	(11)	MOLECULE TYPE: CDNA
55	(11)	MODECORE TIPE: CDNA
	(iii)	HYPOTHETICAL: NO

	(iv) ANTI-SENSE: NO	
5	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2481585	
	(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 2481582	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:	
	GAATTCTTGG ACTATGGGGG CCGGGAACAG GCGATCTTGA GCCGGGTGTC CGGGGTCTCA	60
15	GGGACTGTCA CGTGTTCCCG AGGTGCTTCT AAAACCCTGG CGGCTCCGGA GCCCCTCCTT	120
	CCCACCACCG CCTCCAGGTC CTGCTCCTGC CGCCACCGCT TCCATCTGGA GCAGAAGCGA	180
20	CCGCGCTCAG CCACGTACCC CGGAGTCCAG GCACCCGCAG CGGCCGGGGC ATCCCGAGGA	240
25	TTTTAGT ATG GAG TTT AAG CTT GAG GAG CAT TTT AAC AAG ACA TTT GTC Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val 1 5 10	289
23	ACA GAG AAC AAT ACA GCT GCT GCT CGG AAT GCA GCC TTC CCT GCC TGG Thr Glu Asn Asn Thr Ala Ala Ala Arg Asn Ala Ala Phe Pro Ala Trp 15 20 25 30	337
30	GAG GAC TAC AGA GGC AGC GTA GAC GAT TTA CAA TAC TTT CTG ATT GGG Glu Asp Tyr Arg Gly Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly 35 40 45	385
35	CTC TAT ACA TTC GTA AGT CTT CTT GGC TTT ATG GGC AAT CTA CCT ATT Leu Tyr Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Pro Ile 50 55 60	433
40	TTA ATG GCT GTT ATG AAA AAG CGC AAT CAG AAG ACT ACA GTG AAC TTT Leu Met Ala Val Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe 65 70 75	481
	CTC ATA GGC AAC CTG GCC TTC TCC GAC ATC TTG GTC GTC CTG TTT TGC Leu Ile Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys 80 85 90	529
45	TCC CCT TTC ACC CTG ACC TCT GTC TTG TTG GAT CAG TGG ATG TTT GGC Ser Pro Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly 100 105 110	577
50	AAA GCC ATG TGC CAT ATC ATG CCG TTC CTT CAA TGT GTG TCA GTT CTG Lys Ala Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu 115 120 125	625
	GTT TCA ACT CTG ATT TTA ATA TCA ATT GCC ATT GTC AGG TAT CAT ATG	673

55 Val Ser Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met

											GCA Ala						721
5											GCC						769
10											GAG Glu 185						817
15											TCA Ser						865
20											CTA Leu						913
	Pro	Leu	Val 225	Сув	Leu	Thr	Val	Ser 230	His	Thr	AGC Ser	Val	Суз 235	Arg	Ser	Ile	961
25	Ser	Сув 240	Gly	Leu	Ser	His	Lys 245	Glu	Asn	Arg	CTC Leu	Glu 250	Glu	Asn	Glu	Met	1009
30	_			_		_					AGC Ser 265			_	_		1057
35	Thr	Pro	Ser	Thr	Gln 275	Lys	Trp	Ser	Tyr	Ser 280	TTC Phe	Ile	Arg	Lys	His 285	Arg	1105
40	Arg	Arg	Tyr	Ser 290	Lys	Lys	Thr	Ala	Сув 295	Val	TTA Leu	Pro	Ala	Pro 300	Ala	Gly	1153
45		Ser	Gln 305	Gly	Lys	His	Leu	Ala 310	Val	Pro	GAA Glu	Asn	Pro 315	Ala	Ser	Val	1201
45	Arg	Ser 320	Gln	Leu	Ser	Pro	Ser 325	Ser	Lys	Val	ATT Ile	Pro 330	Gly	Val	Pro	Ile	1249
50											GAT Asp 345						1297
55	_										AGA Arg						1345

	TAC	AGA	CTG	ACC	ATA	CTG	ATA	CTC	GTG	TTC	GCC	GTT	AGC	TGG	ATG	CCA		1393
	Tyr	Arg	Leu	Thr 370	Ile	Leu	Ile	Leu	375	Pne	Ala	var	ser	380	Mec	Pro		
5	CTC	CAC	GTC	TTC	CAC	GTG	GTG	ACT	GAC	TTC	AAT	GAT	AAC	TTG	ATT	TCC		1441
	Leu	His	Val 385	Phe	His	Val	Val	390	Asp	Pne	Asn	Asp	395	Leu	116	ser		
			CAT	TTC				TAC										1489
	Asn	_	His	Phe	Lys	Leu		Tyr	Сув	Ile	Суз	His 410	Leu	Leu	Gly	Met		
10		400					405					410						
				TGT														1537
		Ser	Суз	Cys	Leu	Asn 420	Pro	Ile	Leu	Tyr	Gly 425	Phe	Leu	Asn	Asn	Gly 430		
15	415					420					723					130		
				GAC														1585
	Ile	Lys	Ala	Asp	Leu 435	Arg	Ala	Leu	Ile	H18	Cys	Leu	His	Met	Ser	*		
20	TTC	rcrc'	rgt	GCAC	CAAA	GA G	AGAA	GAAA	GT	egta.	ATTG	ACA	CATA	ATT '	TATA	CAGAA	3	1645
	TAT	rctg	GAT (CTGA	ATGC	CA G	rrcg'	raat(C TAC	CGTA	AGAT	CAT	CTTC	ATG '	TTAT	YTATAA	3	1705
26	GTT	AATT	CAA '	TCAG'	TTGT	GC A	GAGT	CAAT	3 TC	CATC!	TAAT	ACA	ATTT	CAT	GTGT	IGAAG'	r	1765
25	AGT.	TAC	ATT .	ATTT	TCCA'	rt t	ratg'	TCAT:	r gg:	raat:	AAGT	TGA	GTGA:	TAC	TCTG	rggtt	r	1825
										ammai			* * ma	77 X FT	~ » » ~	ייי אייי אייי		1885
	AGT	JTAA	AAG .	ATAT.	AGCT.	AT C	CAAA'	I'I'GI'	r AC	31 T.G.	LACA	AAA	AAIG	IMI	GAAG	TGACA.	н.	1003
30	GTT	GTCC	CAA .	AGAG	CATT	TA A	CTAC	AGAT	TA	AGGA.	TTTA	CTA'	TTAT	CTG	GGTA	TCTTC	A.	1945
	mmm		mmc	3 G3 G	camm	com m	አ አ (ማ አ)	***************************************	in its	ממייי	አአርም	אכא	יממממ	ידי מידי	ጥሮ አ አ	AAGTC	D.	2005
	1111	CIAI	110	ACAG	GCII	Ç1 1.	наса		1 11	GIAA	nu I	ACA	neses.	171		10101		2005
	GAA	CTCT.	ATT	ACAG.	ATGT.	AT G	CATA	AAAG.	A TG	ATTA	TAAT	TTT	GTAG	GAG	AAAG	ATCTG	С	2065
35	ጥር	ידיידי מידי	ልርጥ	GNNG	ል ጥጥር፥	ርጥ A	יימממ	тстс	A GT	TTAA	CCCG	GCT	GTCC	TAC	TACT	AATAT	т	2125
	TAA	TTTT	TCA	ATA	TGAA	AA G	GTTT	CAGA	T TT	TGTT	TAGA	TTT	ATAT	CAC	ATTA	AACAC	T	2185
40	GTC	AAAT	AAA	GGCT	GTTT	TT A	TATG	CATC	G TT	GATG	TTCC	AAA	ATGT	GAA	GTCT	aaatg	G	2245
	TGT	CTGT	ATT	TCCA	ATTA	TT A	ATAA	ACTT	C TA	AGAT	CATT	TTT	AAAA	GTC	TGTA	GATGG	T	2305
	ATG	GATA	GCT	AGTT	GTTT	GT T	AATA	TAAA	G TA	AAAG	TAGA	TAG	CTGA	TTT	ATGT	TGTAC	С	2365
45																		2427
	TAT	GTCG	TAT	GTAT	ATTA	GG T	ATCG	TGTT	G TC	TCAC	TAAA	GTG	AAAG	CAA	ACGA	AAAA	A	2425
	AAA	AAAA	AAA	AAAA	AAAA	AA A	AAAA	AAAA	A AA	аааа	AAAA	AAA	AAAA	AAA	AAAA	AA		2481

(2) INFORMATION FOR SEQ ID NO:2:

50

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 445 amino acids
- 55 (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

		•								_						
5	Met 1		Phe	Lys	Leu 5	Glu	Glu	His	Phe	Asn 10	Lys	Thr	Phe	Val	Thr 15	Glu
	Asn	Asn	Thr	Ala 20	Ala	Ala	Arg	Asn	Ala 25	Ala	Phe	Pro	Ala	Trp 30	Glu	Asp
10	Tyr	Arg	Gly 35		Val	Asp	Asp	Leu 40		Tyr	Phe	Leu	Ile 45		Leu	Tyr
15	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	Asn	Leu 60	Pro	Ile	Leu	Met
	Ala 65	Val	Met	Lys	Lys	Arg 70	Asn	Gln	Lys	Thr	Thr 75	Val	Asn	Phe	Leu	Ile 80
20	Gly	Asn	Leu	Ala	Phe 85	Ser	Asp	Ile	Leu	Val 90	Val	Leu	Phe	Суз	Ser 95	Pro
	Phe	Thr	Leu	Thr 100	Ser	Val	Leu	Leu	Asp 105	Gln	Trp	Met	Phe	Gly 110	Lys	Ala
25	Met	Cys	His 115	Ile	Met	Pro	Phe	Leu 120	Gln	Суѕ	Val	Ser	Val 125	Leu	Val	Ser
30	Thr	Leu 130	Ile	Leu	Ile	Ser	11e 135	Ala	Ile	Val	Arg	Tyr 140	His	Met	Ile	Lys
	His 145	Pro	Ile	Ser	Asn	Asn 150	Leu	Thr	Ala	Asn	His 155	Gly	Tyr	Phe	Leu	Ile 160
35	Ala	Thr	Val	Trp	Thr 165	Leu	Gly	Phe	Ala	Ile 170	Cys	Ser	Pro	Leu	Pro 175	Val
	Phe	His	Ser	Leu 180	Val	Glu	Leu	ГÀа	Glu 185	Thr	Phe	Gly	Ser	Ala 190	Leu	Leu
40	Ser	Ser	Lys 195	Tyr	Leu	Cys	Val	Glu 200	ser	Trp	Pro	Ser	Asp 205	Ser	Tyr	Arg
45	Ile	Ala 210	Phe	Thr	Ile	Ser	Leu 215	Leu	Leu	Val	Gln	Tyr 220	Ile	Leu	Pro	Leu
	Val 225	Сув	Leu	Thr	Val	Ser 230	His	Thr	Ser	Val	Cys 235	Arg	Ser	Ile	Ser	Суз 240
50	Gly	Leu	Ser	His	Lys 245	Glu	Asn	Arg	Leu	Glu 250	Glu	Asn	Glu	Met	Ile 255	Asn
	Leu	Thr	Leu	Gln 260	Pro	Ser	Lys	Lys	Ser 265	Arg	Asn	Gln	Ala	Lys 270	Thr	Pro
55	Ser	Thr	Gln 275	Lys	Trp	Ser	Tyr	Ser 280	Phe	Ile	Arg	Lys	His 285	Arg	Arg	Arg

	Tyr	Ser 290	Lys	Lys	Thr	Ala	Cys 295	Val	Leu	Pro	Ala	Pro 300	Ala	Gly	Pro	Ser
5	Gln 305	Gly	Lys	His	Leu	Ala 310	Val	Pro	Glu	Asn	Pro 315	Ala	Ser	Val	Arg	Ser 320
	Gln	Leu	Ser	Pro	Ser 325	Ser	Lys	Val	Ile	Pro 330	Gly	Val	Pro	Ile	Cys 335	Phe
10	Glu	Val	Lys	Pro 340	Glu	Glu	Ser	Ser	Asp 345	Ala	His	Glu	Met	Arg 350	Val	Lys
	Arg	Ser	11e 355	Thr	Arg	Ile	Lys	360 Lys	Arg	Ser	Arg	Ser	Val 365	Phe	Tyr	Arg
15	Leu	Thr 370	Ile	Leu	Ile	Leu	Val 375	Phe	Ala	Val	Ser	Trp 380	Met	Pro	Leu	His
20	Val 385	Phe	His	Val	Val	Thr 390	Asp	Phe	Asn	Asp	Asn 395	Leu	Ile	Ser	Asn	Arg 400
	His	Phe	Lys	Leu	Val 405	Tyr	Сув	Ile	Cys	His 410	Leu	Leu	Gly	Met	Met 415	Ser
25	Сув	ayɔ	Leu	Asn 420	Pro	Ile	Leu	Tyr	Gly 425	Phe	Leu	Asn	Asn	Gly 430	Ile	Lys
30	Ala	Asp	Leu 435	Arg	Ala	Leu	Ile	His 440	Сув	Leu	His	Met	Ser 445	*		
	(2) INFORMATION FOR SEQ ID NO:3:															
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2604 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear															
40				ECUI				ł.								
		(iii) (iv)		roih: CI-se			VO									
45						. 140										
		(1x)	(2	ATURE A) NA B) LO	ME/I			170	8							
50		(ix)	(2	ATURI A) NI B) LO	ME/I		_									

GAATTCTTGG ACTATGGGGG CCGGGAACAG GCGATCTTGA GCCGGGTGTC CGGGGTCTCA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

55

	GGG	actg:	ICA	CGTG	TTCC	CG A	GGTG	CTTC	T AA	AACC	CTGG	CGG	CTCC	GGA	GCCC	CTCCTT	120
	ccc	ACCA	CCG	CCTC	CAGG	TC C	TGCT	CCTG	C CG	CCAC	CGCT	TCC	ATCT	GGA	GCAG	AAGCGA	180
5	CCG	CGCT	CAG	CCAC	GTAC	cc c	GGAG	TCCA	G GC	ACCC	GCAG	CGG	CCGG	GGC	ATCC	CGAGCT	240
	GGC	CATA	CAC	CGGG	AGAC.	AG C	TGTG	CCCT	T GG	GTTT	GCAA	GGT	GGCT	TGG	aagt	CAACTG	300
	CCA	GTAG	GAA .	ATAG	CCAT	CC A	CACA	CCTG	A GT	TCCA	AGGG	GGA	AGAA	AGA	GATT	CTTATC	360
10	TGA'	TTTT	AGT .	ATG	GAG '	TTT .	AAG	CTT	GAG (GAG	CAT	TTT .	AAC	AAG .	ACA '	TTT	409
	Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe 1 5 10																
15												GCA					457
	val	15	GIU	ASII	ABII	THE	20	Ата	ALA	Arg	Asn	Ala 25	Ala	Pue	PIO	Ala	
20										_		CAA					505
20	Trp 30		qaA	Tyr	Arg	Gly 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	Ile 45	
												ATG					553
25	Gly	Leu	Tyr	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	Asn	Leu 60	Leu	
	ATT	TTA	ATG	GCT	GTT	ATG	AAA	AAG	CGC	AAT	CAG	AAG	ACT	ACA	GTG	AAC	601
	_											Lys					
30	ւկական	רייוירי	מידמ		አልሮ	CTG	GCC	ምምር		GNC	አ ምርግ	TTG	C.T.C.		caca	diction	649
	_		Ile					Phe				Leu	Val				043
35	maa	maa	80					85					90				
33		Ser										GAT Asp					697
		95					100					105					
40												CAA Gln					745
	110															125	
												ATT Ile					793
45					130		200		541	135	7,14	116	var	Arg	140	1173	
												GCA					841
	Mec	TIE	гув	145	PIO	TTE	ser	ASI	ASN 150	ren	Thr	Ala	Asn	155	GIA	Tyr	
50												GCC					889
	Phe	Leu	Ile 160	Ala	Thr	Val	Trp	Thr 165	Leu	Gly	Phe	Ala	Ile 170	Суз	Ser	Pro	
55												GAG					937
	neu	175	val	rue.	nıs	ser	180	val	GIU	Leu	гла	Glu 185	Inr	ьие	СΤÀ	ser	

							TCA Ser			985
5							CTA Leu			1033
10							AGC Ser			1081
							CTC Leu			1129
15							AGC Ser 265			1177
20							TTC Phe			1225
25							TTA Leu			1273
30							GAA Glu			1321
35							ATT Ile			1369
40							GAT Asp 345			1417
40	 	 	 				AGA Arg			1465
45							GCC Ala			1513
50							AAT Asn			1561
55					Tyr		TGT Cys	Leu		1609

	ATG ATG TCC TGT TGT CTA AAT CCG ATC CTA TAT GGT TTC CTT AAT AAT Met Met Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn 415 420 425	1657
5	GGT ATC AAA GCA GAC TTG AGA GCC CTT ATC CAC TGC CTA CAC ATG TCA Gly Ile Lys Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser 430 435 440 445	1705
10	TGA TTCTCTCTGT GCACCAAAGA GAGAAGAAAC GTGGTAATTG ACACATAATT *	1758
	TATACAGAAG TATTCTGGAT CTGAATGCCA GTTCGTAATC TACGTAAGAT CATCTTCATG	1818
15	TTATAATATG GTTAATTCAA TCAGTTGTGC AGAGTCAATG TCCATCTAAT ACAATTTCAT	1878
	GTGTTGAAGT AGTTTACATT ATTTTCCATT TTATGTCATT GGTAATAAGT TGAGTGATAC	1938
20	TCTGTGGTTT AGTGTAAAAG ATATAGCTAT CCAAATTGTT ACGTTGTACA AAAAATGTAT	1998
20	GAAGTGACAA GTTGTCCCAA AGAGCATTTA ACTACAGATT TAAGGAATTT CTATTATCTG	2058
	GGTATCTTCA TTTCTATTTC ACAGGCTTCT TAACATTTTT TTGTAAAAGT ACAAAAATAT	2118
25	TCAAAAGTCA GAACTCTATT ACAGATGTAT GCATAAAAGA TGATTATAAT TTTGTAGGAG	2178
	AAAGATCTGC TCCTATTAGT GAAGATTGGT AAAATTGTCA GTTTAACCCG GCTGTCCTAC	2238
30	TACTAATATT TAATTTTTCA AATATGAAAA GGTTTCAGAT TTTGTTTAGA TTTATATCAC	2298
50	ATTAAACACT GTCAAATAAA GGCTGTTTTT ATATGCATCG TTGATGTTCC AAAATGTGAA	2358
	GTCTAAATGG TGTCTGTATT TCCAATTATT AAATAACTTC TAAGATCATT TTTAAAAGTC	2418
35	TGTAGATGGT ATGGATAGCT AGTTGTTTGT TAATATAAAG TAAAAGTAGA TAGCTGATTT	2478
	ATGTTGTACC TATGTCGTAT GTATATTAGG TATCGTGTTG TCTCACTAAA GTGAAAGCAA	2538
40	АССБАВАВАВ ВАВАВАВАВ ВАВАВАВАВ ВАВАВАВАВ ВАВАВАВАВ В	2598
→ 0	ААААА	2604
	(2)	

(2) INFORMATION FOR SEQ ID NO:4:

45

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 445 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

50

- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
- 55 Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val Thr Glu
 1 5 10 15

	Asn	Asn	Thr	Ala 20	Ala	Ala	Arg	Asn	Ala 25	Ala	Phe	Pro	Ala	Trp 30	Glu	Asp
5	Tyr	Arg	Gly 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	Ile 45	Gly	Leu	Tyr
	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	Asn	Leu 60	Leu	Ile	Leu	Met
10	Ala 65	Val	Met	Lys	ГÀЗ	Arg 70	Asn	Gln	Lys	Thr	Thr 75	Val	Asn	Phe	Leu	Ile 80
15	Gly	Asn	Leu	Ala	Phe 85	Ser	Asp	Ile	Leu	Val 90	Val	Leu	Phe	Cys	Ser 95	Pro
	Phe	Thr	Leu	Thr 100	Ser	Val	Leu	Leu	Asp 105	Gln	Trp	Met	Phe	Gly 110	Lys	Ala
20	Met	Сув	His 115	Ile	Met	Pro	Phe	Leu 120	Gln	Сув	Val	Ser	Val 125	Leu	Val	Ser
	Thr	Leu 130	Ile	Leu	Ile	Ser	Ile 135	Ala	Ile	Val	Arg	Tyr 140	His	Met	Ile	Lys
25	His 145	Pro	Ile	Ser	Asn	Asn 150	Leu	Thr	Ala	Asn	His 155	Gly	Tyr	Phe	Leu	Ile 160
30	Ala	Thr	Val	Trp	Thr 165	Leu	Gly	Phe	Ala	Ile 170	Cys	Ser	Pro	Leu	Pro 175	Val
	Phe	His	Ser	Leu 180	Val	Glu	Leu	Lys	Glu 185	Thr	Phe	Gly	Ser	Ala 190	Leu	Leu
35	Ser	Ser	Lys 195	Tyr	Leu	Cya	Val	Glu 200	Ser	Trp	Pro	Ser	Asp 205	Ser	Tyr	Arg
40	Ile	Ala 210	Phe	Thr	Ile	Ser	Leu 215	Leu	Leu	Val	Gln	Tyr 220	Ile	Leu	Pro	Leu
	Val 225		Leu	Thr	Val	Ser 230	His	Thr	Ser	Val	Cys 235		Ser	Ile	Ser	Cys 240
45	Gly	Leu	Ser	His	Lys 2 4 5	Glu	Asn	Arg	Leu	Glu 250		. Asn	Glu	Met	Ile 255	
	Leu	Thr	Leu	Gln 260		Ser	Lys	Lys	Ser 265		Asn	Gln	Ala	Lys 270		Pro
50	Ser	Thr	Gln 275		Trp	Ser	Tyr	Ser 280		lle	Arg	. Lys	His 285		Arg	Arg
55	Tyr	Ser 290		Lys	Thr	Ala	Cys 295		. Leu	Pro	Ala	300		. Gly	Pro	Ser
<i>J</i> .	Gln 305	_	Lys	His	Leu	Ala 310		Pro	Glu	Asn	9rc 315		ser	· Val	. Arg	320

	Gln	Leu	Ser	Pro	Ser 325	Ser	Lys	Val	Ile	Pro 330	_	Val	Pro	Ile	Суз 335		
5	Glu	Val	Lys	Pro 340	Glu	Glu	Ser	Ser	Asp 345	Ala	His	Glu	Met	Arg 350	Val	Lys	
	Arg	Ser	Ile 355	Thr	Arg	Ile	Lys	143 360	Arg	Ser	Arg	Ser	Val 365	Phe	Tyr	Arg	
10	Leu	Thr 370	Ile	Leu	Ile	Leu	Val 375	Phe	Ala	Val	Ser	Trp 380	Met	Pro	Leu	His	
15	385		His			390					395					400	
			Lys		405					410					415		
20			Leu	420					425					430	Ile	Lys	
	Ala	АЗР	Leu 435	Arg	Ala	ren	TTE	440	Cys	Leu	HIS	met	445	•			
25	(2)	INFO	ORMAT	NOI	FOR	SEQ	ID N	10:5:									
30		(i)	(E	L) LE () TY () SI	ngth Pe : Rani	IARAC I: 13 nucl EDNE OGY:	38 l eic SS:	ase acid sing	pair l	s							
		(ii)	MOL	ECUL	E TY	PE:	genc	mic	DNA								
35	(iii)	HYP	OTHE	TICA	L: N	O										
		(iv)	ANT	T-SE	NSE :	NO											
40		(ix)		.) NA	ME/K	EY:		.338									
45		(ix)		AN (ME/K	EY:			ide								
		(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NC	:5:						
50	ATG Met																48
	AAT 2														GAT		96
55	Asn (Asn	Thr .	Ala . 20	Ala	Thr .	Arg .	Asn	Ser 25	Asp	Phe	Pro	Val	Trp 30	Авр	qaA	

				AGT Ser													144
5				AGT Ser													192
10				AAA Lys												_	240
15				GCC Ala													288
20				ACG Thr 100													336
	Met	Сув	His 115	ATT Ile	Met	Pro	Phe	Leu 120	Gln	Сув	Val	Ser	Val 125	Leu	Val	Ser	384
25	Thr	Leu 130	Ile	TTA Leu	Ile	Ser	Ile 135	Ala	Ile	Val	Arg	Tyr 140	His	Met	Ile	Lys	432
30				TCT Ser													480
35				TGG Trp													528
40				CTT Leu 180													576
	Ser	Ser	Arg 195	TAT Tyr	Leu	Сув	Val	Glu 200	Ser	Trp	Pro	Ser	Asp 205	Ser	Tyr	Arg	624
45				ACT Thr													672
50				ACT Thr													720
55				AAC Asn													768

											CCT Pro					816
5											AAA Lys					864
10											GCT Ala					912
15											AAC Asn 315					960
											CCA Pro					1008
20											GTT Val					1056
25											TCT Ser					1104
30											GTT Val					1152
35											GAC Asp 395					1200
											CAT His					1248
40											TTT Phe					1296
45	AAA Lys	GCT Ala	GAT Asp 435	TTA Leu	GTG Val	TCC Ser	CTT Leu	ATA Ile 440	CAC His	тст Сув	CTT Leu	CAT His	ATG Met 445	TAA *		1338

50 (2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 445 amino acids

(B) TYPE: amino acid

55 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

5	Met 1	Asp	Leu	Glu	Leu 5	Asp	Glu	Tyr	Tyr	Asn 10	Lys	Thr	Leu	Ala	Thr 15	Glu
J	Asn	Asn	Thr	Ala 20	Ala	Thr	Arg	Asn	Ser 25	Asp	Phe	Pro	Val	Trp 30	Asp	Asp
10	Tyr	Lys	Ser 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	Ile 45	Gly	Leu	Tyr
	Thr	Ph e 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	Asn	Leu 60	Leu	Ile	Leu	Met
15	Ala 65	Leu	Met	Lys	Lys	Arg 70	Asn	Gln	Lys	Thr	Thr 75	Val	Asn	Phe	Leu	Ile 80
20	Gly	Asn	Leu	Ala	Phe 85	Ser	Asp	Ile	Leu	Val 90	Val	Leu	Phe	Сув	Ser 95	Pro
	Phe	Thr	Leu	Thr 100	Ser	Val	Leu	Leu	Asp 105	Gln	Trp	Met	Phe	Gly 110	Lys	Val
25	Met	Сув	His 115	Ile	Met	Pro	Phe	Leu 120	Gln	Суз	Val	Ser	Val 125	Leu	Val	Ser
	Thr	Leu 130	Ile	Leu	Ile	Ser	Ile 135	Ala	Ile	Val	Arg	Tyr 140	His	Met	Ile	Lys
30	His 145	Pro	Ile	Ser	Asn	Asn 150	Leu	Thr	Ala	Asn	His 155	Gly	Tyr	Phe	Leu	Ile 160
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	Phe	His	Ser	Leu 180	Val	Glu	Leu	Gln	Glu 185	Thr	Phe	Gly	Ser	Ala 190	Leu	Leu
40	Ser	Ser	Arg 195	Tyr	Leu	Cys	Val	Glu 200	Ser	Trp	Pro	Ser	Asp 205	Ser	Tyr	Arg
	Ile	Ala 210	Phe	Thr	Ile	Ser	Leu 215	Leu	Leu	Val	Gln	Tyr 220	Ile	Leu	Pro	Leu
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	Leu	Thr	Leu	His 260	Pro	Ser	Lys	Гуз	Ser 265		Pro	Gln	Val	Lys 270		Ser
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	Tyr	Ser 290		Lys	Thr	Ala	Cys 295	Val	Leu	Pro	Ala	Pro 300	Glu	Arg	Pro	Ser	
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	Ser	Gln	Leu	Ser	Ser 325	Ser	Ser	Lys	Phe	Ile 330	Pro	Gly	Val	Pro	Thr 335	Cys	
10	Phe	Glu	Ile	Lуя 340	Pro	Glu	Glu	Asn	Ser 345	Asp	Val	His	Glu	Leu 350	Arg	Val	
15	Lys	Arg	Ser 355	Val	Thr	Arg	Ile	Lys 360	Lys	Arg	Ser	Arg	Ser 365	Val	Phe	Tyr	
	Arg	Leu 370	Thr	Ile	Leu	Ile	Leu 375	Val	Phe	Ala	Val	Ser 380	Trp	Met	Pro	Leu	
20	His 385	Leu	Phe	His	Val	Val 390	Thr	Asp	Phe	Asn	Asp 395	Asn	Leu	Ile	Ser	Asn 400	
		His			405					410				-	415		
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	Lys	Ala	Asp 435	Leu	Val	Ser	Leu	Ile 440	His	Cys	Leu	His	Met 445	*			
30	(2)	INFO	RMAT	rion	FOR	SEQ	ID N	iO : 7 :									
35		(i)	() (E	A) LE 3) TY C) SI	TE CHENGTH TPE: TRANK	: 20 nucl EDNE	bas eic SS:	e pa acid sing	irs								
40		(ii)	MOL	ECUL	E TY	PE:	othe	r nu	clei	c ac	id						
40																	
		(xi)	SEC	UENC	E DE	SCRI	PTIC	N: S	EQ I	D NO	:7:						
45	TAGG	GAAC	CT G	GCCI	CCTC	:C											20
	(2)	INFO															
50		(i)	(A (B (C	LE TY	E CH NGTH PE: RAND POLO	: 20 nucl EDNE	bas eic SS:	e pa acid sing	irs								
55		(ii)	MOL	ECUL	E TY	PE:	othe	r nu	clei	c ac	iđ						

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TCAGAGGGCC ATGACTCAAC

20

We claim:

1. An isolated nucleic acid encoding a neuropeptide Y receptor comprising a molecule having a nucleotide sequence substantially the same as SEQ ID NO 1, SEQ ID NO 3, or SEQ ID NO 5.

- 2. A homogenous composition of the neuropeptide Y receptor comprising a molecule having an amino acid sequence substantially the same as SEQ ID NO 2, SEQ ID NO 4, or SEQ ID NO 6.
- 3. A vector comprising the nucleic acid according to claim 1.
- 4. A vector according to claim 3 adapted for expression in a cell further comprising regulatory elements necessary for expression of the nucleic acid operatively linked to the nucleic acid to enable expression of the nucleic acid.
- 5. A vector according to claim 4 wherein the cell is a mammalian cell.
- 6. A vector according to claim 5 wherein the cell is a human 293 cell.
- 7. A vector according to claim 4 that is a plasmid.
- 8. A vector according to claim 7 wherein the plasmid is the pBluescript plasmid.
- 9. A vector according to claim 7 wherein the plasmid is the pcDNA3 plasmid.
- 10. A vector according to claim 3 which is self-replicating.
- 11. A cell transformed with the nucleic acid according to claim 1 that expresses the nucleic acid.
- 12. A cell according to claim 11 that is a bacterium cell, an insect cell, or a yeast cell.
- 13. A cell according to claim 11 that is a mammalian cell.
- 14. A cell according to claim 13 that is a human 293 cell.

15. A nucleic acid probe comprising a nucleic acid complementary to the nucleic acid according to claim 1.

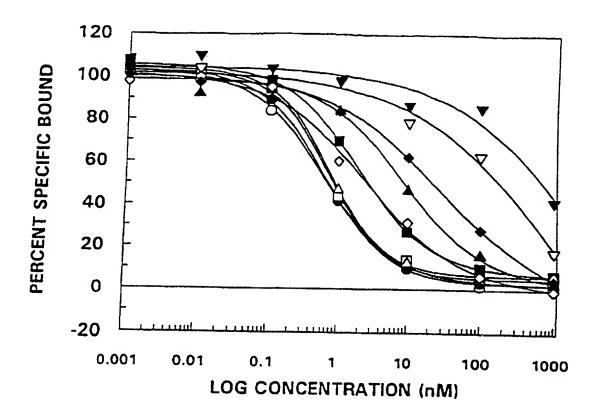
- 16. An antisense oligonucleotide having a sequence complementary to the nucleic acid according to claim 1 and that inhibits expression of the nucleic acid.
- 17. A membrane or membrane preparation comprising a membrane or portion thereof of a cell expressing a nucleic acid having a nucleotide sequence substantially the same as SEQ ID NO 1, SEQ ID NO 3, or SEQ ID NO 5.
- 18. A membrane or membrane preparation according to claim 17 wherein the cell is a mammalian cell.
- 19. An antibody or fragment thereof that is immunologically reactive to a mammalian Y5 receptor.
- 20. An antibody or fragment according to claim 19 thereof wherein the mammalian Y5 receptor has an amino acid sequence substantially the same as one chosen from the group consisting of SEQ ID NO 2, SEQ ID NO 4, and SEQ ID NO 6.
- 21. An antibody or fragment thereof according to claim 19 that is a monoclonal antibody.
- 22. An antibody or fragment thereof according to claim 20 that is a monoclonal antibody.
- 23. A cell line producing an antibody according to claim 19.
- 24. A cell line producing an antibody according to claim 20.
- 25. A cell line producing an antibody according to claim 21.
- 26. A cell line producing an antibody according to claim 22.
- 27. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 19.
- 28. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 20.

29. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 21.

- 30. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 22.
- 31. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 11 and recovering the receptor expressed by the cell.
- 32. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 12 and recovering the receptor expressed by the cell.
- 33. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 13 and recovering the receptor expressed by the cell.
- 34. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 14 and recovering the receptor expressed by the cell.
- 35. A method of identifying a neuropeptide Y agonist or antagonist comprising contacting a potential agonist or antagonist molecule with a membrane or membrane preparation according to claim 17.
- 36. A method of identifying a neuropeptide Y agonist or antagonist comprising contacting a potential agonist or antagonist molecule with a membrane or membrane preparation according to claim 18.
- 37. A neuropeptide Y antagonist comprising a compound identified according to claim 36.
- 38. A method of suppressing the appetite of a mammal comprising administering to the mammal an appetite suppressing amount of a neuropeptide Y antagonist according to claim 37.
- 39. A method of suppressing the appetite of a mammal according to claim 38 wherein the amount of antagonist is from about 0.01 to about 100 mg/kg.
- 40. A pharmaceutical composition comprising an effective appetite suppressing amount of an antagonist according to claim 37 together with a pharmaceutically acceptable carrier.

41. A neuropeptide Y agonist comprising a compound identified according to claim 36.

- 42. A method of stimulating the appetite of a mammal comprising administering to the mammal an appetite stimulating amount of a neuropeptide Y agonist according to claim 41.
- 43. A method of stimulating the appetite of a mammal according to claim 42 wherein the amount of agonist is from about 0.01 to about 100 mg/kg.
- 44. A pharmaceutical composition comprising an effective appetite stimulating amount of an agonist according to claim 41 together with a pharmaceutically acceptable carrier.
- 45. A non-human transgenic mammal that expresses the nucleic acid having a sequence substantially the same one chosen from the group consisting of SEQ ID NO 1, SEQ ID NO 3, and SEQ ID NO 5.



○ NPY
 ● PYY
 □ (LPINPY)
 ■ (3-36)NPY
 △ (2-36)NPY
 △ (13-36)NPY
 ▽ (18-36)NPY
 ▽ (20-36)NPY
 ◇ (3-36)(LP)NPY
 ◆ (10-36)NPY

Fig. 1

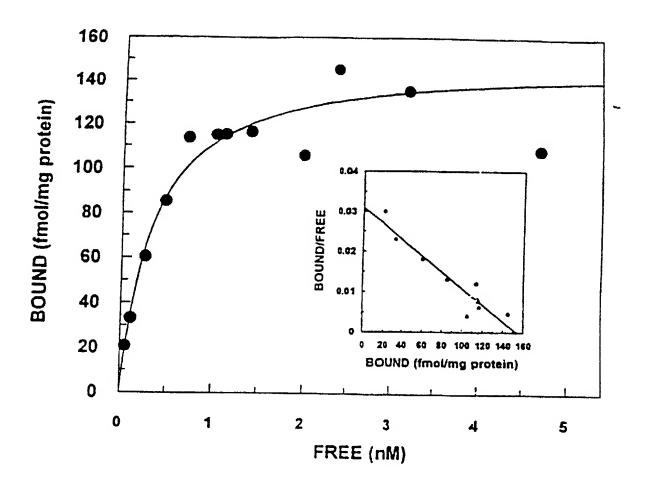


Fig. 2



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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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US

(71) Applicant: BAYER CORPORATION [US/US]; 100 Bayer Road, Pittsburgh, PA 15205 (US).

(72) Inventors: HU, Yinghe; 17 Falcon Crest Drive, North Haven, CT 06473 (US). McCALEB, Michael, L.; 447 Bartlett Drive, Madison, CT 06443 (US). BLOOMQUIST, Brian, T.; 405 Stevenson Road, New Haven, CT 06515 (US). FLORES-RIVEROS, Jaime, R.; 27 Carmel Court, Madison, CT 06443 (US). CORNFIELD, Linda, J.; 3 Hidden Brook Road, Hamden, CT 06518 (US).

(74) Agents: GREENMAN, Jeffrey, M. et al.; Bayer Corporation, 400 Morgan Lane, West Haven, CT 06516 (US).

(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, IP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).

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With international search report.

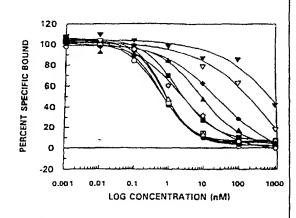
Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

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(54) Title: NEUROPEPTIDE Y RECEPTOR Y5 AND NUCLEIC ACID SEQUENCE

(57) Abstract

The present invention provides novel NPY/PYY receptor proteins and the nucleic acid sequence encoding them. The invention is directed to the isolation, characterization, and pharmacological use of these receptors and nucleic acids. In particular, this invention provides human and rat NPY/PYY receptors (which we call the NPY Y5 receptor) and nucleic acids. Also provided are recombinant expression constructs useful for transfecting cells and expressing the protein in vitro and in vivo. The invention further provides methods for detecting expression levels of the protein as well as methods for screening for receptor antagonists and agonists to be used for the treatment of obesity or anorexia, respectively.





- PY'
- □ ILPINPY
- (3-36)NPY
- △ (2-36)NPY
- ▲ (13-36)NPY
- ▼ (20-36)NPY
- ♦ (10-36)NPY

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Inter. nal Application No PCT/US 97/05781

				PC	1/05 97/05/81
A. CLASS IPC 6	IFICATION OF SUBJEC C12N15/12 A61K38/02	C07K14/705 C12Q1/68	C12N5/10 C07K16/28		A01K67/027
According	to International Patent Clas	sification (IPC) or to be	th national classification	n and IPC	
B. FIELDS	S SEARCHED				
Minimum o	coumentation searched (cl CO7K A61K	assification system follo	wed by classification sy	mbols)	
Documenta	tion searched other than m	nimum documentation	to the extent that such o	locuments are included	in the fields searched
Electronic	data hase consulted during (he international search (name of data base and	, where practical, search	s terms used)
C. DOCUN	MENTS CONSIDERED TO) BE RELEVANT			
Category *	Citation of document, wi	th indication, where app	ropnate, of the relevan	t passages	Relevant to claim No.
Х	NEUROPEPTID TARGETING O	: "SUBTYPES E Y: IMPLICAT F THERAPEUTIC	IONS FOR THE		37,38, 41,42,44
A	see page 55 see page 55		3; table 1 3 - paragrap		1,2
	see page 55		-/		
V Fur	her documents are listed in	the combination of box	c lv	Peters family member	ers are listed in annex.
			с. Х		
'A' docume consider a filing of the consider of the consideration of the	ent which may throw doubt is cited to establish the pub n or other special reason (a ent referring to an oral disc	te of the art which is not vance or after the internation; s on priority claim(s) or lication date of another s specified) lostire, use, exhibition o	.X. q	or priority date and not intend to understand the privention occurrent of particular reannot be considered nonvolve an inventive step occurrent of particular reannot be considered to: annot be considered to: focurrent is combined we	after the international filing date in conflict with the application but rinciple or theory underlying the clevance; the claimed invention well or cannot be considered to when the document is taken alone clevance; the claimed invention involve an inventive step when the inth one or more other such docubeing obvious to a person skilled
later th	nan the priority date claime	d	ui.	ocument member of the	same patent family
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Category *	WAHLESTEDT C ET AL: "NEUROPEPTIDE Y-RELATED PEPTIDES AND THEIR RECEPTORS-ARE THE RECEPTORS POTENTIAL THERAPEUTIC DRUG TARGETS?"	37,38, 41,42
X	WAHLESTEDT C ET AL: "NEUROPEPTIDE Y-RELATED PEPTIDES AND THEIR RECEPTORS-ARE THE RECEPTORS POTENTIAL THERAPEUTIC DRUG	37,38,
X	Y-RELATED PEPTIDES AND THEIR RECEPTORS-ARE THE RECEPTORS POTENTIAL THERAPEUTIC DRUG	
	ANNUAL REVIEW OF PHARMACOLOGY AND TOXICOLOGY, vol. 32, 1993, pages 309-352, XP000612055 see page 320, paragraph 2 see page 327, paragraph 4 - page 328, line 6 see page 338, paragraph 3 see page 335, paragraph 3 - paragraph 5 see page 340, paragraph 4 see page 341, paragraph 6 - page 342, line 3	
X	KAZUHIKO TATEMOTO ET AL: "SYNTHESIS OF RECEPTOR ANTAGONISTS OF NEUROPEPTIDE Y" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 89, no. 4, pages 1174-1178, XP000259387 see the whole document	35-37,41
(WO 93 12139 A (GARVAN INST MED RES ;PRINCE OF WALES MEDICAL RESEAR (AU)) 24 June 1993 see the whole document	37-39
`	WO 93 09227 A (GARVAN INST MED RES) 13 May 1993 see claims	1,3-14, 17,18, 31-37,41
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, X	GERALD, C. ET AL.: "A receptor subtype involved in neuropeptide-Y-induced food intake" NATURE, vol. 382, no. 6587, 11 July 1996, LONDON GB, pages 168-171, XP000612078 see the whole document	2
,х	WO 96 16542 A (SYNAPTIC PHARMA CORP) 6 June 1996 see the whole document	1-45

5 2

Inter anal Application No
PCT/US 97/05781

Category Citation of	JMENTS CONSIDERED TO BE RELEVANT document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
	document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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indu a no ENDO vol. page	EA, D. ET AL.: "Neuropeptide Y ced feeding in the rat is mediated by vel receptor" CRINOLOGY, 138, no. 1, January 1997, s 196-202, XP002040613 the whole document	1
HEIN RIGO Sequ	7 20821 A (CIBA GEIGY AG ;RUEEGER RICH (CH); SCHMIDLIN TIBUR (CH); LL) 12 June 1997 ence listing claims	1-45
expr rece JOUR (MIC vol.	BERG, D.H. ET AL.: "Cloning and ession of a novel neuropeptide Y ptor" NAL OF BIOLOGICAL CHEMISTRY ROFILMS), 271, no. 28, 12 July 1996, MD US, s 16435-16438, XP002040614 page Y	1
anal rece JOUR (MIC vol. page	, P.M. ET AL.: "Molecular genetic ysis of a human neuropeptide Y ptor" NAL OF BIOLOGICAL CHEMISTRY ROFILMS), 272, no. 6, 7 February 1997, MD US, s 3622-3627, XP002040615 page 3622, column 2, line 12 - line 27	1
stru rece coor GENO vol. page	OG, H. ET AL.: "Overlapping gene cture of the human neuropeptide Y ptor subtypes Y1 and Y5 suggests dinate transcription regulation" MICS, 41, no. 3, 1 May 1997, s 315-319, XP002040616 the whole document	

5

rnational application No. PCT/US 97/05781

INTERNATIONAL SEARCH REPORT

Box 1	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	rnational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: see FURTHER INFORMATION sheet PCT/ISA/210
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3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inter	national Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all earchable claims.
2	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report overs only those claims for which fees were paid, specifically claims Nos.:
4 r	to required additional search fees were timely paid by the applicant. Consequently, this International Search Report is estricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark o	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM	PCT/ISA/	210
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Remark:

As far as claims 38,39,42 and 43 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

information on patent family members

Inter anal Application No
PCT/US 97/05781

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